

VitriLife[®] Technology Platform

Industrial Scale Production & Mucosal Delivery of
Thermostable Vaccines and Other Biopharmaceuticals

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Markets We Serve

Cold chain requirements due to the thermal instability of freeze-dried (F-D) vaccines and many other biologicals limit their stockpile potential, distribution, delivery, and greatly increase costs. Most current vaccines are formulated for parenteral delivery, requiring trained medical personnel, sterile water, needles and syringes. There is an urgent need for thermostable vaccines that can be self-administered.

VitriLife® technology platform addresses a need for better ambient temperature (AT) stabilization and delivery of vaccines and other fragile biologicals with higher yields, at higher storage temperatures, and lower cost of production.



**Biopharmaceuticals
Including Vaccines**



**Animal Health &
Nutrition**

**VitriLife®
Markets**



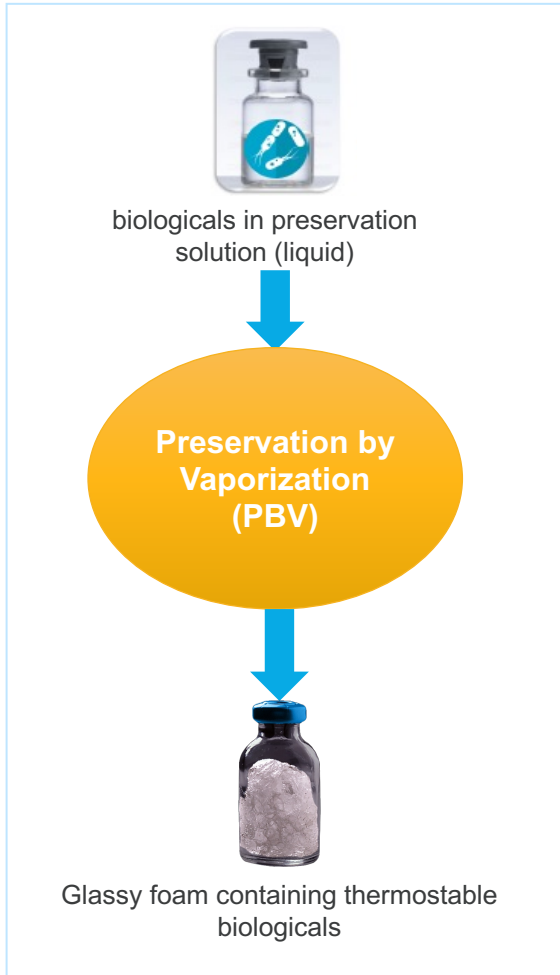
Agricultural Biologicals



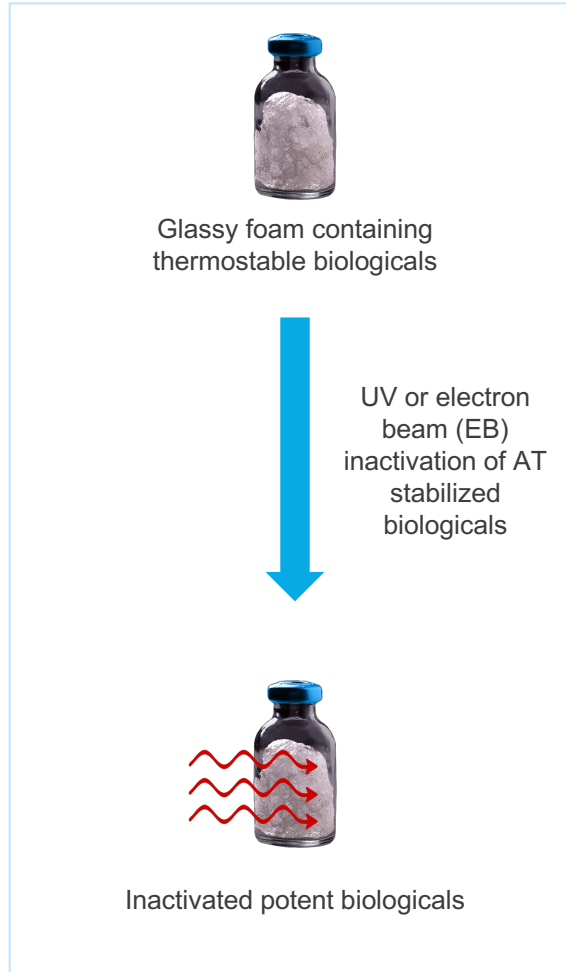
Human Microbiome

Three Pillars of VitriLife[®] Technology Platform

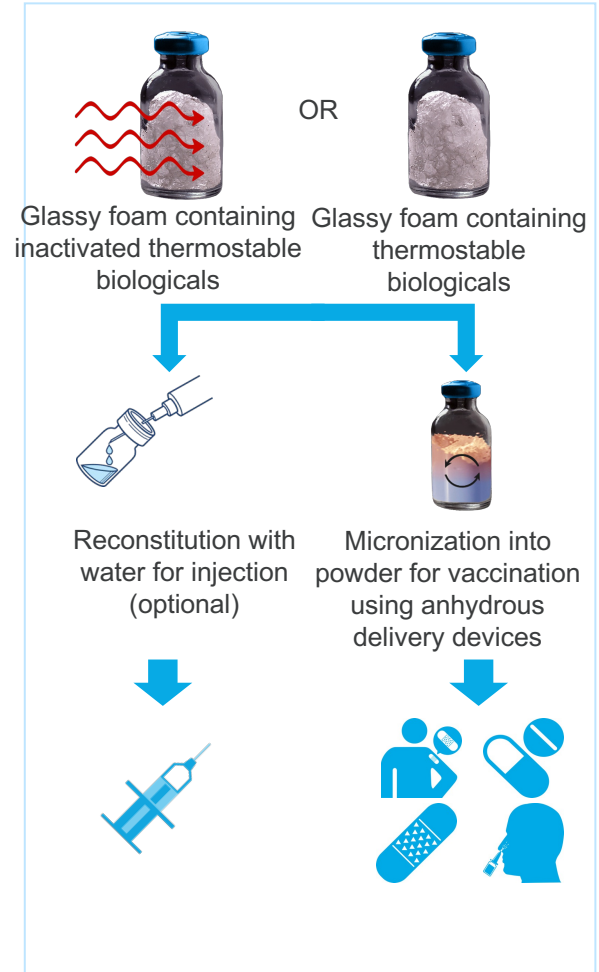
AT Stabilization



Inactivation (Optional)



Delivery



Key Benefits of VitriLife® Technologies

Stabilization

- Cornerstone of UST's *VitriLife*® platform is its proprietary Preservation by Vaporization (PBV) industrial scale vacuum drying technology, which preserves biologicals by boiling from a partially frozen (slush) state. PBV enables substantially higher AT stabilization of vaccines, microbiome and other heat-labile biotherapeutics than can be achieved using conventional lyophilization.
- PBV is several times quicker and cheaper than freeze-drying, can be executed using standard lyophilizers, and is suitable for GMP under FDA/EMA approval.
- Biologicals stabilized using PBV technology remain stable at high ambient temperatures, eliminating a need for a cold chain during storage and distribution.
- Shorter drying time vs. freeze drying (minutes to hours vs. hours to days), removes bottlenecks associated with lyophilization manufacturing.
- Lower losses in vaccine activity or potency vs. lyophilization.
- PBV enables novel and more efficient vaccine inactivation and delivery technologies.

Inactivation

- PBV stabilized products can be treated with UV light or electron-beam (EB) irradiation in a dry state at AT, allowing for controllable, targeted damage, primarily to nucleic acids.
- UV or EB inactivation of microorganisms after PBV has significant advantages vs. heat or chemical inactivation, which damage membranes and biomolecules.
- These inactivation technologies also enable accelerated and cost-effective development and production of vaccines utilizing wild-type pathogens, which can address urgent vaccination needs against emerging diseases.

Delivery

- Thermostable *VitriLife*® vaccines and other biologicals can be micronized with no or minimum damage for subsequent incorporation in needle-free delivery devices (including DPIs, disintegrable tablets or capsules, dissolvable polymeric rods, films, microneedle patches, etc.). These devices enable mucosal (respiratory, oral, intravaginal etc.) and transdermal delivery without reconstitution and minimize the need for trained medical personnel for administration.

VitriLife[®] Stabilization Overview

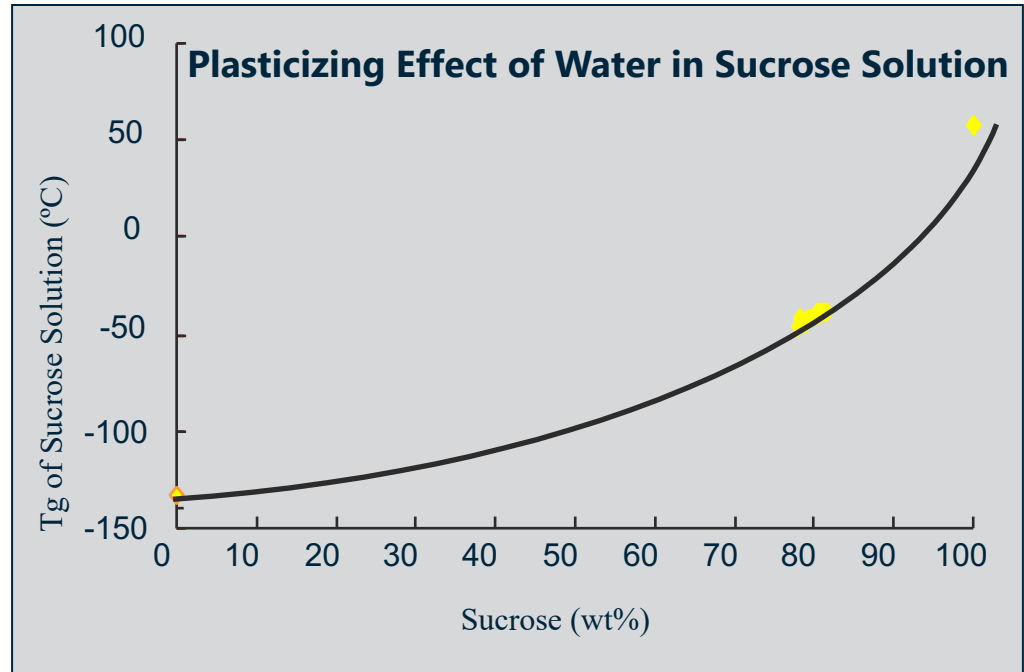
Ambient Temperature (AT) Stability Can be Achieved by Immobilization of biologicals in a Carbohydrate Glass



Insect Preserved in Amber Glass

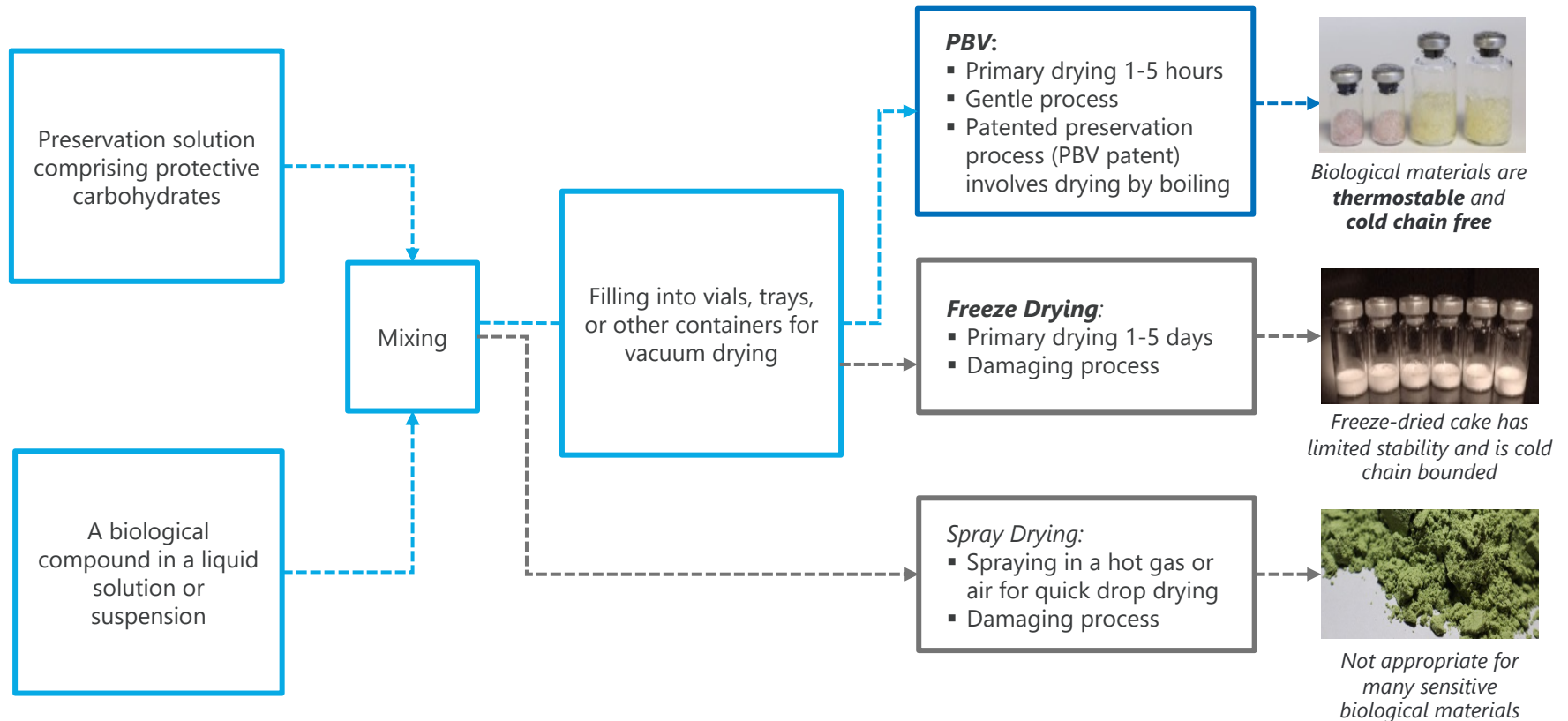


L. acidophilus Preserved in Milled Sugar Glass



Water decreases glass transition temperature (T_g) and should be removed to achieve glass state at ambient temperatures.

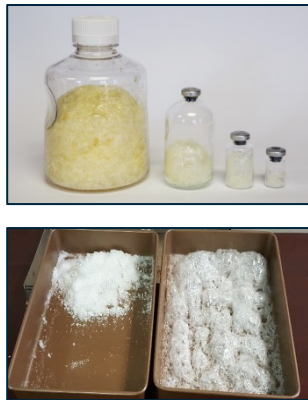
PBV vs. Conventional Freeze-Drying and Spray Drying



VitriLife (PBV) Stabilization is Scalable for Industrial Applications

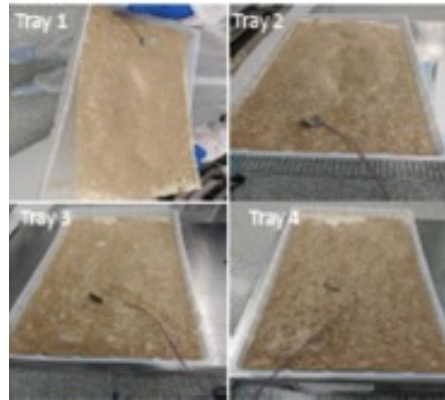
The images below depict the scale-up of *VitriLife*® stabilized dairy cow rumen microbes from a small lab to mass production at one of the largest lyophilization manufacturing facilities.

Laboratory Scale



0.5cm fill height
Batch size: 0.3kg

R&D Scale



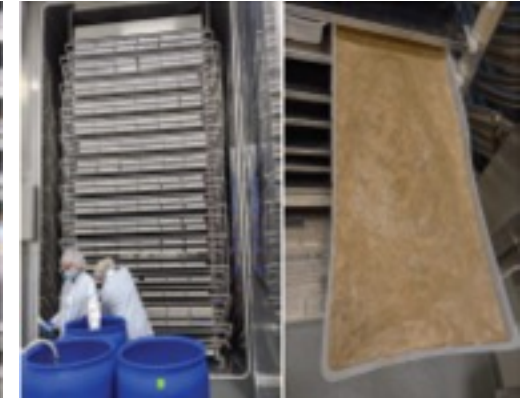
0.5cm to 1.8cm fill height
Batch size: 0.6kg to 3.8kg

Production Scale



1.8cm fill height
Max batch size: 41.8kg
(110 trays x 3.8kg)

Enhanced Production Scale

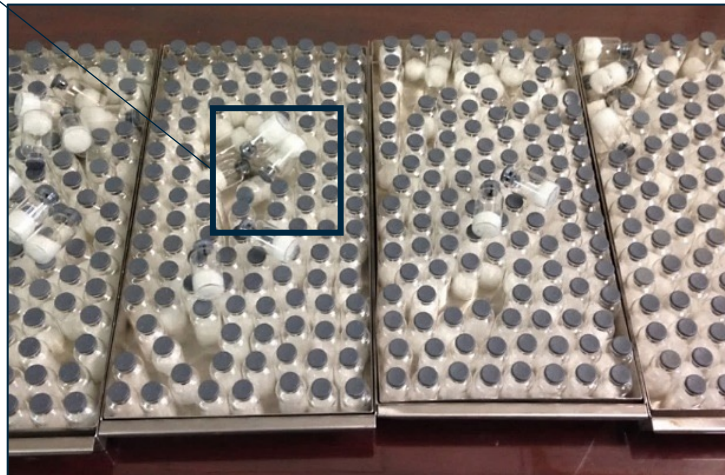


1.8cm fill height
Max batch size: 205.2kg
(540 trays x 3.8kg)

PBV is a Commercially Validated Superior Drying Technology for Industrial Applications at Scale

PBV is a Reproducible and Scalable Process (I)

No Evidence of Significant Variability Across Multiple Trays and within a Single Tray



Bulk and unit dose formats:

- ✓ Vials
- ✓ Trays
- ✓ Bottles
- ✓ Containers
- ✓ Bags
- ✓ Other

Reproducibility Testing

Vials taken from 4 locations on full drying tray:

Glass Transition Temp (T_g)

Group # Group Average (T_g)

| | |
|---|-----------------|
| 1 | 48.18 ± 3.17 °C |
| 2 | 47.08 ± 2.07 °C |
| 3 | 49.48 ± 3.07 °C |
| 4 | 48.65 ± 1.99 °C |

Water Activity (a_w)

Group # Group Average (a_w)

| | |
|---|-----------------|
| 1 | 0.1287 ± 0.0060 |
| 2 | 0.1290 ± 0.0012 |
| 3 | 0.1319 ± 0.0017 |
| 4 | 0.1305 ± 0.0032 |

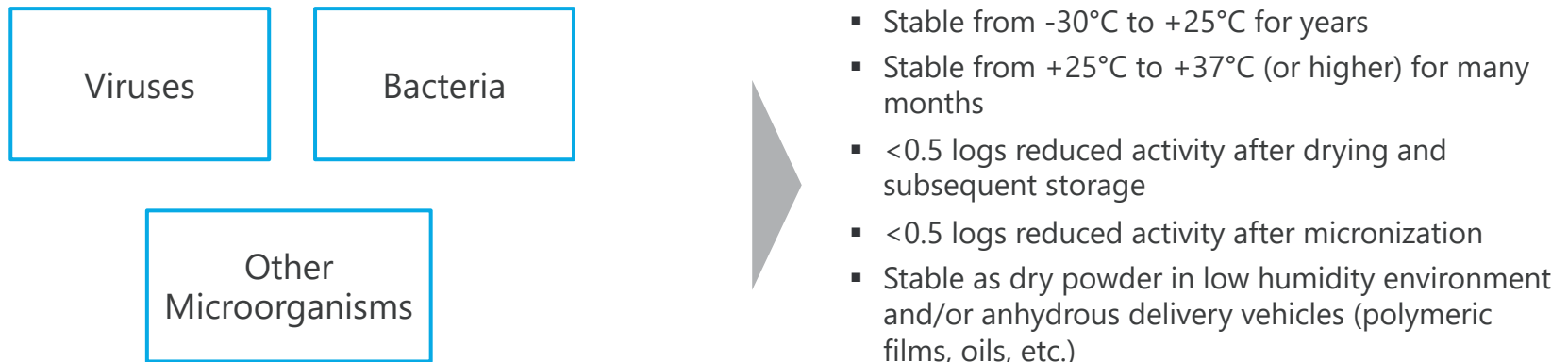
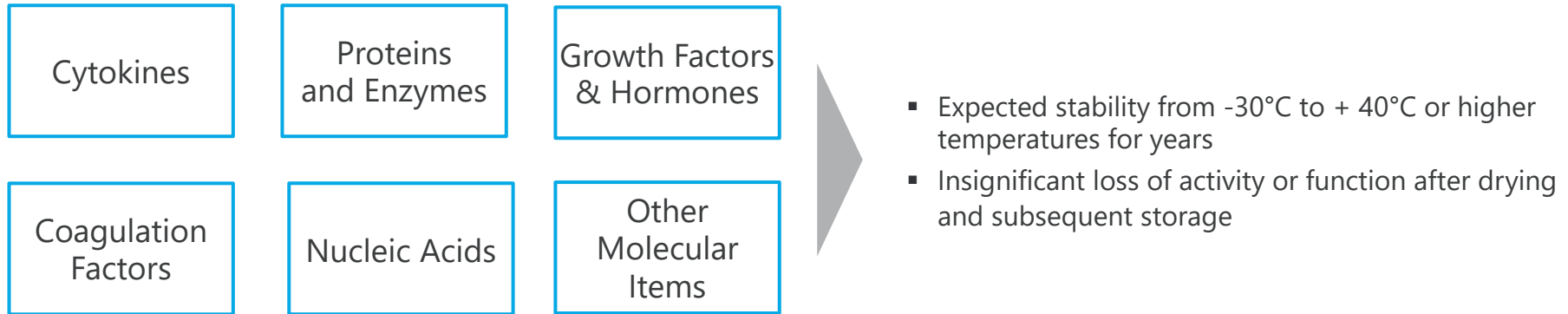
PBV is a Reproducible and Scalable Process (2)

Three vials were evaluated from four corners and from the center of each tray after two independent PBV drying runs. No evidence of significant vial-to-vial variability in *Listeria Monocytogenes* LAV survival, T_g , A_w , nor foam physical appearance were found.

| PBV Run #1 | Location on a tray | Viability after PBV (E9 CFU/mL) | Water activity (A_w) Mean of 3 | T_g (°C) Mean of 3 |
|---------------|--------------------|---------------------------------|------------------------------------|----------------------|
| Before Drying | N/A | 1.96± 0.28 | N/A | N/A |
| 1 | Corner 1 | 1.94 ± 0.08 | 0.127 | 25.1 |
| 1 | Corner 2 | 2.09 ± 0.05 | 0.142 | 25.3 |
| 1 | Corner 3 | 2.25 ± 0.01 | 0.137 | 24.8 |
| 1 | Corner 4 | 1.92 ± 0.16 | 0.118 | 25.5 |
| 1 | Middle | 2.38 ± 0.11 | 0.112 | 25.9 |
| 2 | Corner 1 | 1.82 ± 0.09 | 0.135 | 24.5 |
| 2 | Corner 2 | 1.70 ± 0.09 | 0.136 | 25.6 |
| 2 | Corner 3 | 1.90 ± 0.30 | 0.122 | 25.9 |
| 2 | Corner 4 | 1.69 ± 0.09 | 0.139 | 25.4 |
| 2 | Middle | 2.03 ± 0.06 | 0.126 | 26.1 |

| PBV Run #2 | Location on a tray | Viability after PBV (E9 CFU/mL) | Water activity (A_w) Mean of 3 | T_g (°C) Mean of 3 |
|---------------|--------------------|---------------------------------|------------------------------------|----------------------|
| Before Drying | N/A | 2.75± 0.01 | N/A | N/A |
| 1 | Corner 1 | 1.77 ± 0.14 | 0.127 | 24.4 |
| 1 | Corner 2 | 1.94 ± 0.24 | 0.127 | 24.8 |
| 1 | Corner 3 | 1.70 ± 0.08 | 0.127 | 25.2 |
| 1 | Corner 4 | 1.74 ± 0.05 | 0.129 | 25.8 |
| 1 | Middle | 1.65 ± 0.08 | 0.124 | 24.0 |
| 2 | Corner 1 | 1.65 ± 0.05 | 0.118 | 25.2 |
| 2 | Corner 2 | 1.42 ± 0.11 | 0.117 | 25.2 |
| 2 | Corner 3 | 1.54 ± 0.28 | 0.127 | 24.9 |
| 2 | Corner 4 | 1.42 ± 0.12 | 0.129 | 24.2 |
| 2 | Middle | 2.52 ± 0.14 | 0.123 | 23.2 |

PBV Preserves Viability of Microorganisms and Activity of Biomolecules at High and Low Ambient Temperatures (AT)

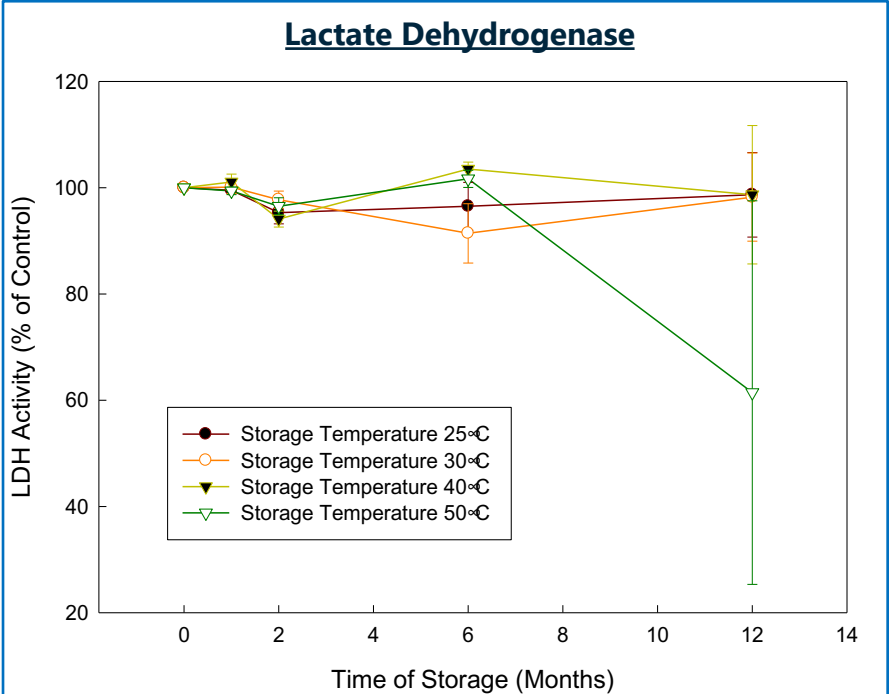
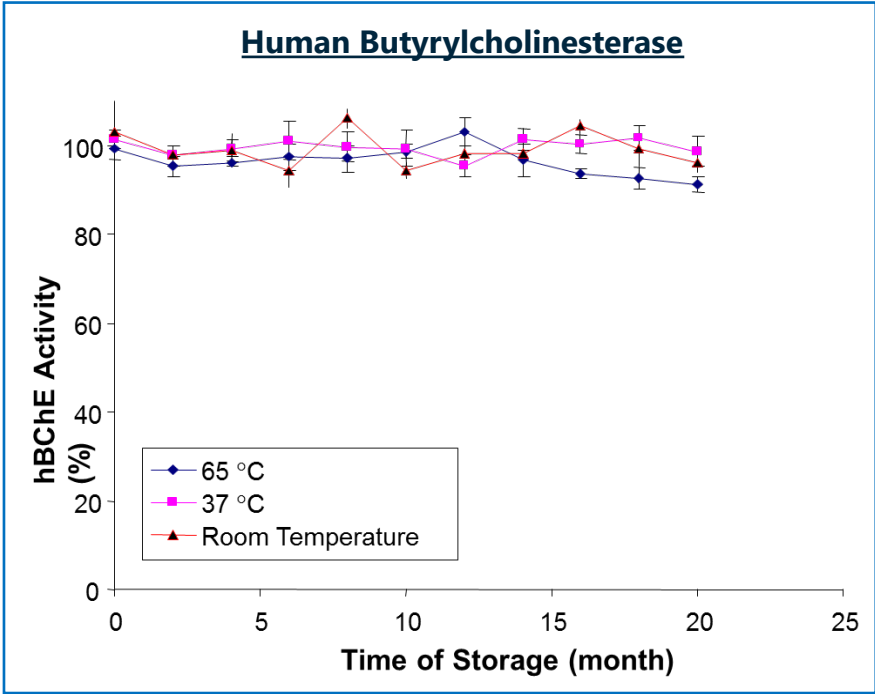


Stability of VitriLife® Proteins / Molecular Items

No Activity Loss after Drying and Subsequent Storage

| Class | Material | Temperature | Storage Time | Activity Loss |
|---------------------------|--------------------------|-------------|--------------|---------------|
| Coagulation Factor | Factor VIII | 40°C | 90 days | No loss |
| Coagulation Factor | Factor VII | 37°C | 40 days | No loss |
| Coagulation Factor | Factor XIII | 37°C | 40 days | No loss |
| Industrial Enzyme | Ice Nucleating Protein | 50°C | 99 days | No loss |
| | | 37°C | 140 days | No loss |
| Enzyme | Thrombine (blood) | 37°C | 40 days | No loss |
| Enzyme (serine proteases) | Urokinase Human Protein | 40°C | 30 days | No loss |
| Enzyme | Luciferase | 37°C | 180 days | No loss |
| Enzyme | β-galactosidase | 37°C | 176 days | No loss |
| Enzyme | Lactate Dehydrogenase | 40°C | 360 days | No loss |
| | | 50°C | 180 days | No loss |
| Enzyme | Isocitrate Dehydrogenase | 50°C | 70 days | No loss |
| Antibiotic | Amphotericin | 40°C | 30 days | No loss |

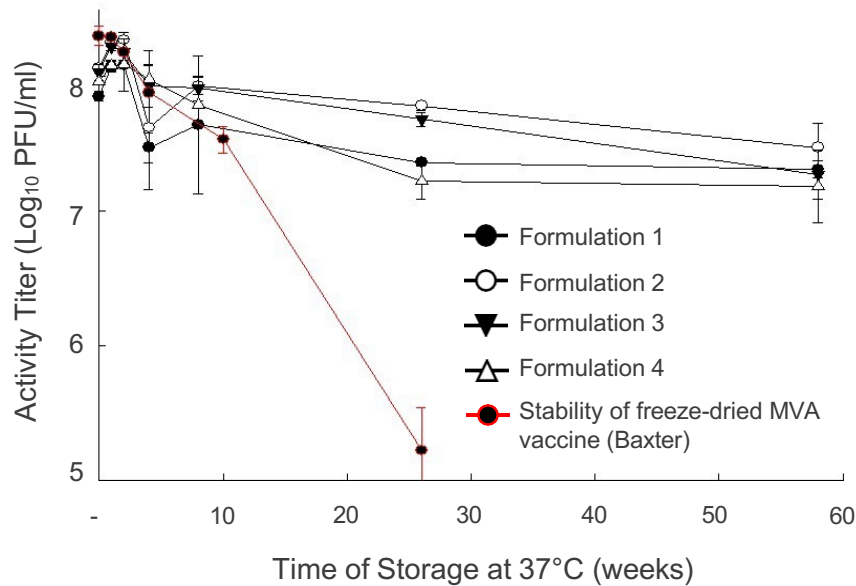
Superior Stability of VitriLife[®] Enzymes at High Ambient Temperatures (AT)



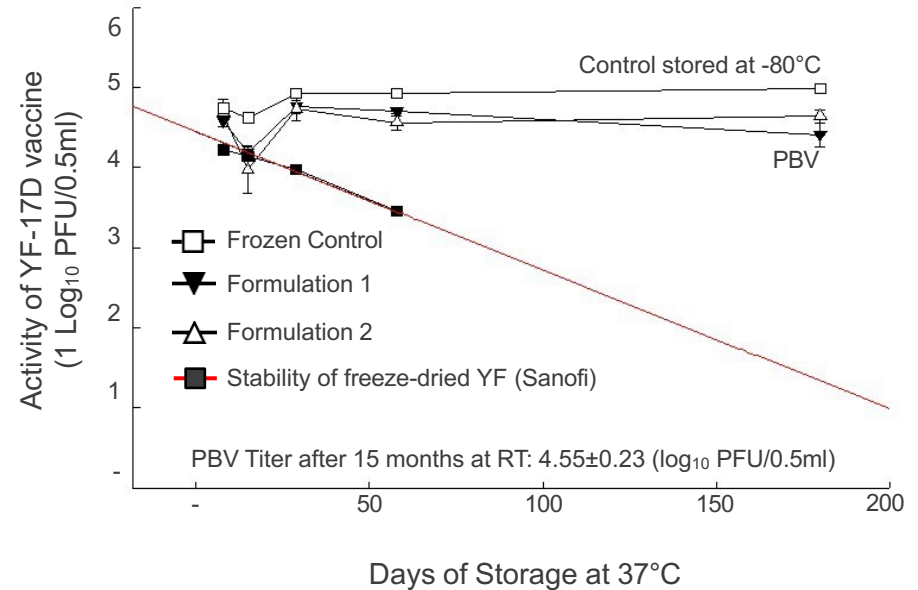
Head-to-Head Stability Comparison of PBV vs. FD LAVs

Commercial FD Vaccines Reconstituted with UST's Preservation Solutions and Stabilized Using PBV

MVA (Smallpox) Vaccine from Baxter



Yellow Fever Vaccine from Sanofi

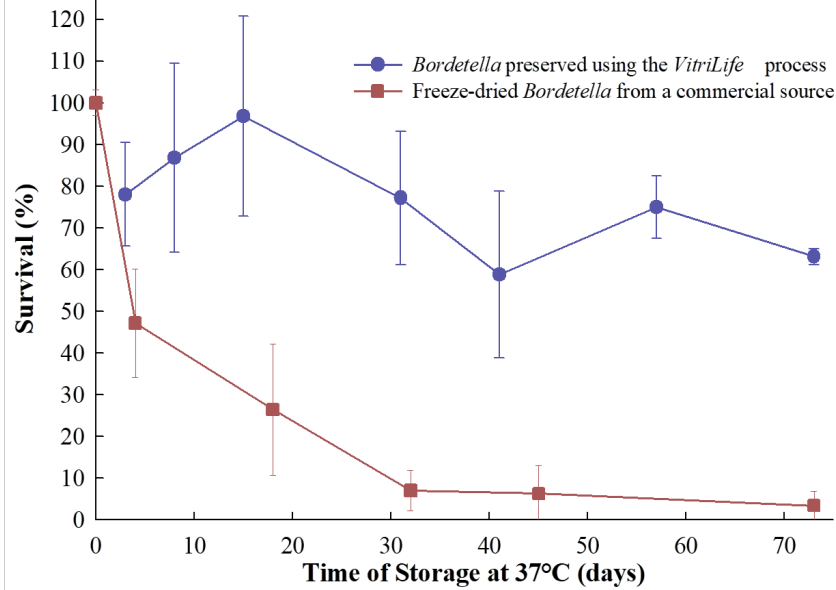


*Additional stability comparison data available for Measles and Bordetella

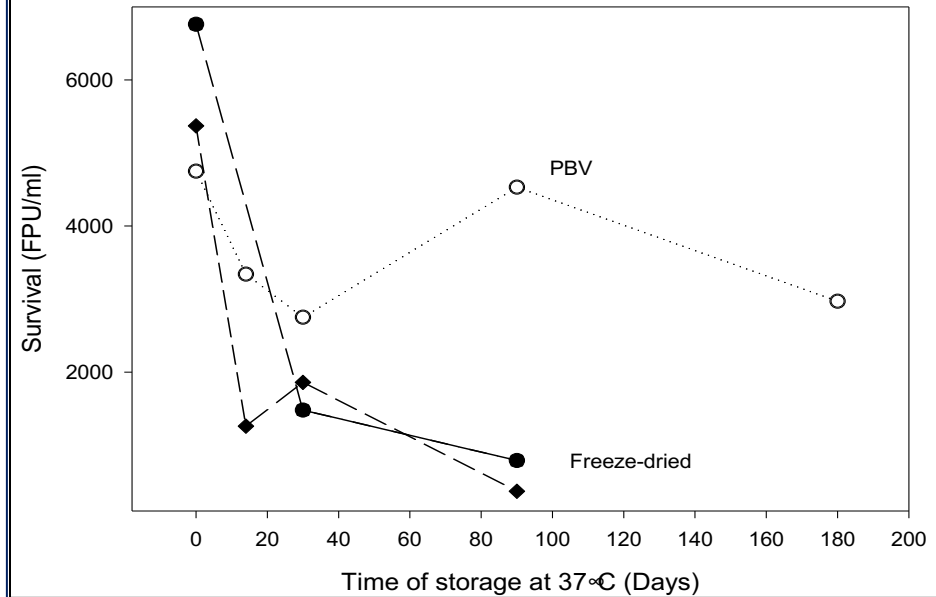
At 37°C, Freeze-Dried LAVs' Activity is Reduced >1,000x when Stored for Six Months, While PBV LAVs Remain Stable

Head-to-Head Stability Comparison of PBV vs. FD LAVs

Bordetella Vaccine



Measles Vaccine from Berna Biotech



Stability of VitriLife® Live Attenuated Vaccines (LAVs)

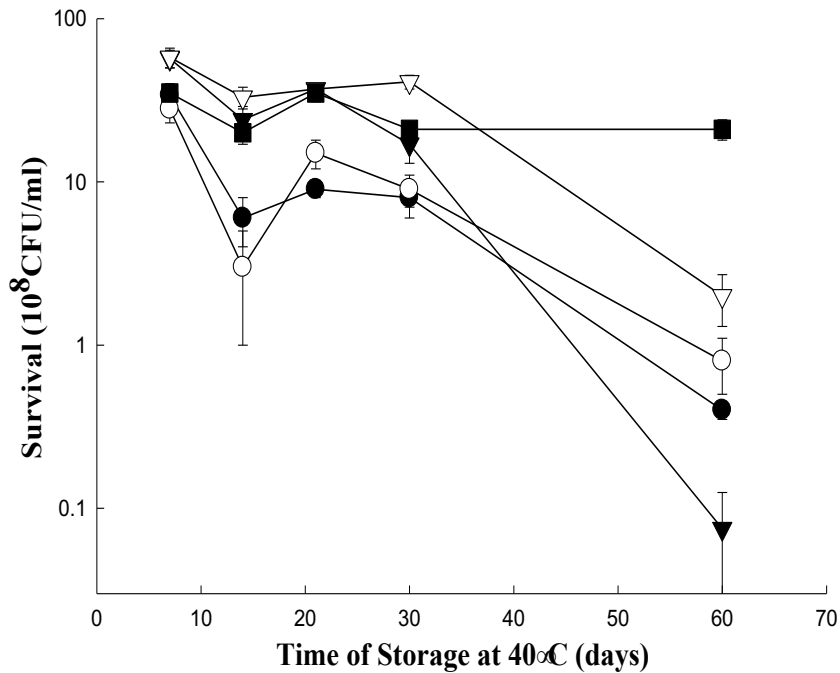
Freeze-Drying Loses > 1 Log and Requires Refrigerated Storage; VitriLife® Loses <0.5 Logs and is Cold-Chain Free

Activity / Viability of VitriLife® Vaccines after Drying and Subsequent Storage

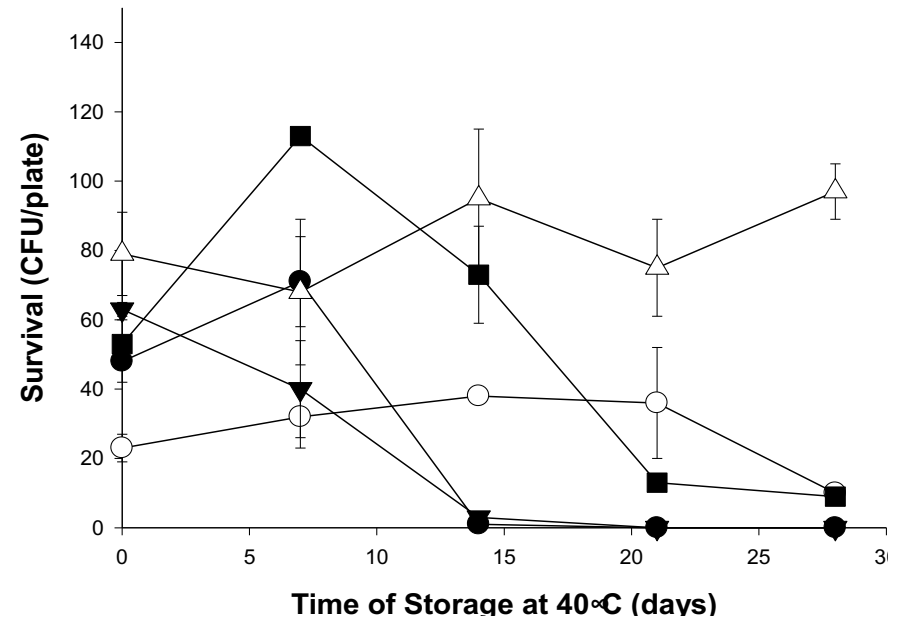
| Live Attenuated Vaccines | 37°C | | 25°C | |
|--------------------------------|--------------|---------------|--------------|---------------|
| | Storage Time | Activity Loss | Storage Time | Activity Loss |
| Rubella | 6 months | 0.5 log | 1 year | 0.5 log |
| Influenza | 3 months | 0.1 log | 1 year | 0 log |
| Rotavirus | 2 years | 0.5 log | 2 years | 0 log |
| Polio Sabin strain | 1.5 months | 0.4 log | 1.5 months | 0 log |
| Modified Vaccinia Ankara (MVA) | 1 year | 0.6 log | 3 years | 0.3 log |
| Yellow Fever | 8 months | 0.2 log | 1 year | 0 log |
| Rabies | 2 months | 0.6 log | 2 years | 0.4 log |
| Measles (from SII) | 4 months | 0.3 log | 1 year | 0.7 log |
| Measles (from CDC) | 2 years | 0.7 log | | |
| Listeria Monocytogenes | 12 months | 0.2 log | | |
| Vegetative Anthrax | 10 months | 0.1 log | | |

Stability of PBV-preserved probiotic bacteria at 40°C depends on composition of preservation solutions (PS)

Stability of *L. reuteri* after PBV with different PS



Stability of *L. rhamnosus* after PBV with different PS



Stability of VitriLife® Gram-Positive Probiotic Bacteria

We found that the membranes of many Lactobacilli and other gram-positive bacteria tend to be desiccation tolerant, and success of their preservation depends on the successful loading of the intracellular space with carbohydrates to protect intracellular proteins, enzymes, nucleic acids and other biologically active molecules during drying.

UST has the expertise and know-how to formulate and optimize preservation solutions and cell loading protocols.

| | <i>L. crispatus</i> | <i>L. rhamnosus</i> | <i>L. jensenii</i> | <i>L. monocytogenes</i> |
|-------------------------|---------------------------|---------------------|--------------------|-------------------------|
| Viability | (x10 ⁹ CFU/mL) | | | |
| Before drying | 9.5 ± 2.8 | 13.9 ± 1.7 | 11.8 ± 1.2 | 32.6 ± 1 |
| After Drying | 7.0 ± 1.2 | 9.3 ± 1.5 | 11.0 ± 1.5 | 17.7 ± 0.7 |
| 3 months at 37°C | 5.2 ± 0.9 | 7.8 ± 0.6 | 11.6 ± 2.0 | |
| 6 months at 37°C | 5.2 ± 1.2 | 4.9 ± 0.7 | 3.1 ± 0.7 | |
| 1 year at 37°C | 3.3 ± 0.5 | 7.6 ± 0.7 | 4.2 ± 0.3 | 11.7 ± 1.6 |
| 1 year at 25°C | 5.5 ± 1.9 | 11.3 ± 1.0 | 8.6 ± 1.5 | |
| 2 years at 25°C | 6.0 ± 0.7 | 8.9 ± 0.3 | 7.4 ± 0.9 | |
| 3 years at 25°C | | 5.1 ± 0.8 | 7.9 ± 1.0 | |

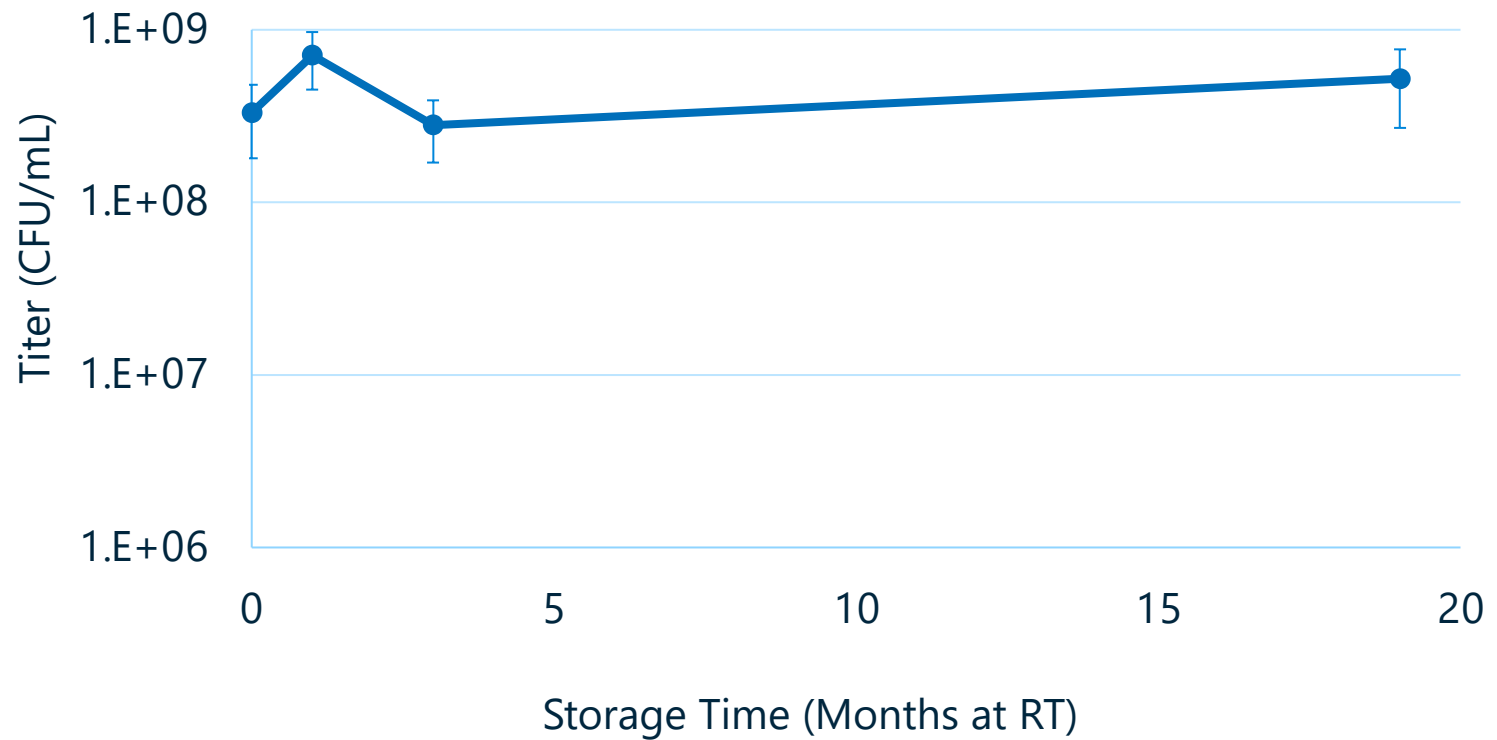
Stability of VitriLife® Gram-Negative Probiotic Bacteria

It is difficult to stabilize gram-negative bacteria and other microorganisms with more fragile membranes. In addition to loading these cells with protective carbohydrates, fortification of the extracellular membranes may be required to achieve thermostable products.

In many cases, the fortification can be achieved using polymeric protectants that reversibly cross-link and/or preferentially absorb to the membrane surface. We identified several polymers that can be used to fortify these membranes and help produce thermostable gram-negative bacteria.

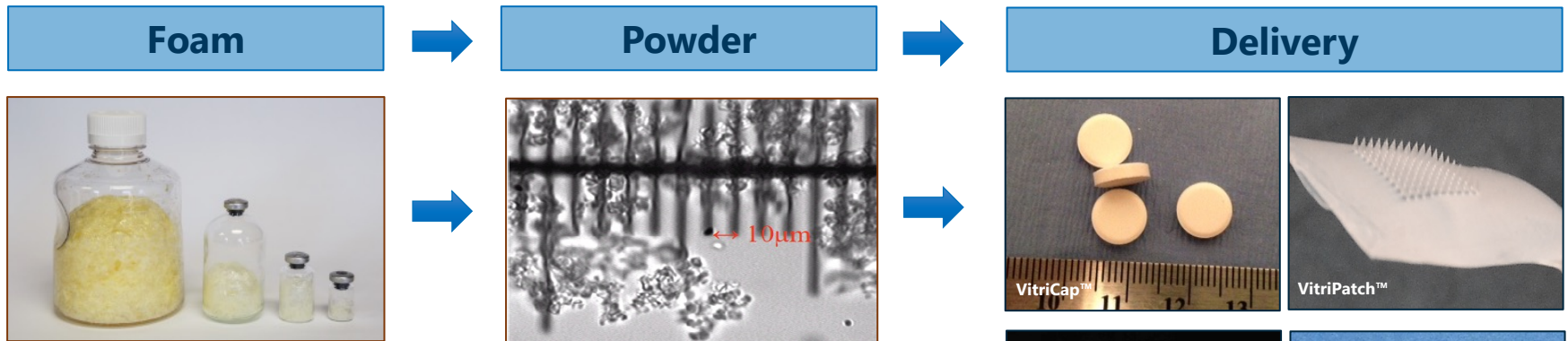
| Viability | E. coli M17 (x106 CFU/mL) | K. variicola (x108 CFU/mL) |
|-------------------------|--------------------------------------|---------------------------------------|
| Before Drying | 465 ± 39 | 140 ± 50 |
| After Drying | 460 ± 90 | 99 ± 8 |
| 1-month at RT | 303 ± 65 | |
| 1-month at 37°C | 204 ± 50 | 61 ± 9 |
| 3 months at RT | 287 ± 32 | |
| 3 months at 37°C | 162 ± 30 | 49 ± 5 |

Long-term Stability of Probiotic E. coli M17 at Room Temperature (RT)



VitriLife[®] Delivery Overview

Needle-Free Delivery without Reconstitution

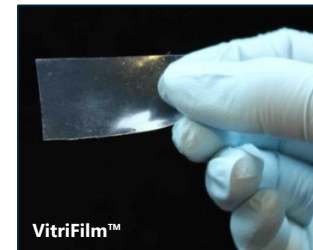


Routes:

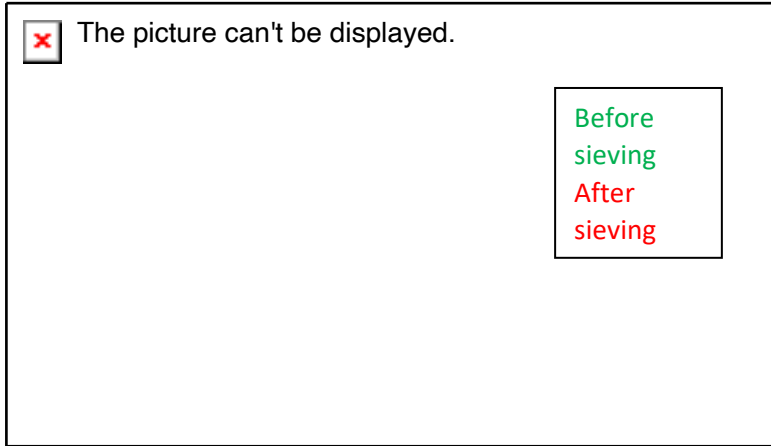
- Oral (intestinal, sublingual, buccal)
- Respiratory/intranasal
- Transdermal
- Vaginal
- Rectal

Devices:

- DPLs
- Capsules/tablets
- Oil suspensions
- Dissolvable films
- Microneedle patches
- Ointment/creams
- Suppositories



Stability of Micronized *VitriLife*[®] H3N2 LAIV

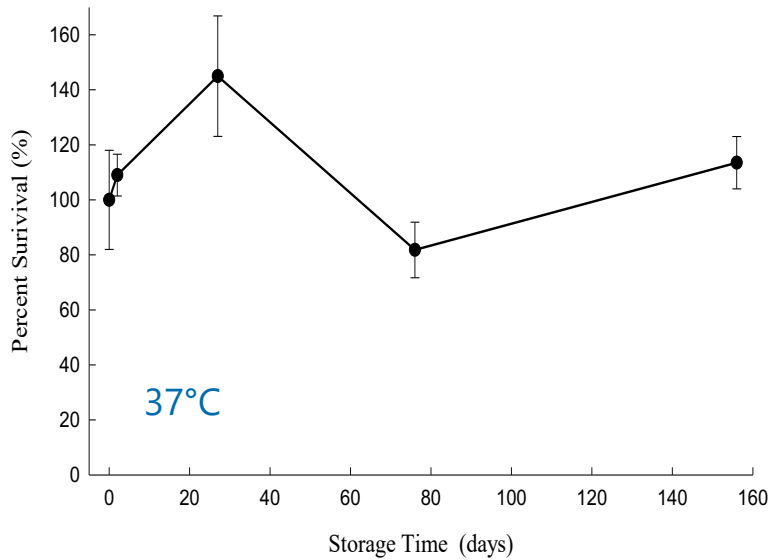


Vaccine was micronized and sieved between 21-63 microns for optimal respiratory delivery. There was minimal activity loss after micronization and good subsequent stability at ATs.

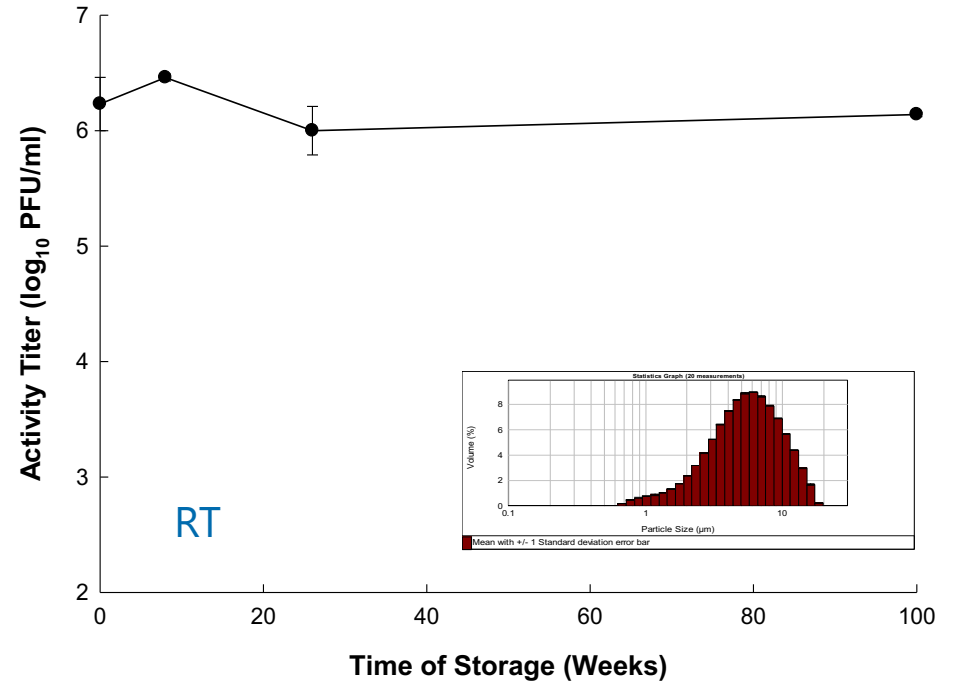
| Initial Yield and Stability of LAIV in final Micronized Powder Format | | | | | |
|---|----------------|------------------|----------------------------------|----------------------------------|------------------------------------|
| Timepoint | Formulation | Overall Log loss | Average (TCID ₅₀ /mL) | Average (TCID ₅₀ /mg) | Deviation (TCID ₅₀ /mg) |
| Initial Yield | Frozen control | N/A | 1.34E+07 | 1.12E+05 | 5.02E+04 |
| | Flu-7 Powder | -0.46 | 4.69E+06 | 3.90E+04 | 8.25E+03 |
| 1 Month Stability | Frozen control | N/A | 1.05E+07 | 8.77E+04 | 3.79E+04 |
| | RT powder | -0.46 | 3.62E+06 | 3.02E+04 | 2.19E+03 |
| | 37°C powder | -0.44 | 3.83E+06 | 3.19E+04 | 1.56E+04 |
| 3 Month Stability | Frozen control | N/A | 1.21E+07 | 2.53E+04 | 0.83E+04 |
| | RT powder | -0.55 | 3.44E+06 | 7.16E+03 | 3.31E+03 |
| | 37°C powder | -0.83 | 1.77E+06 | 3.70E+03 | 0.42E+03 |
| 6 Month Stability | Frozen control | N/A | 4.40E+07 | 9.16E+04 | 2.22E+04 |
| | RT powder | -0.56 | 1.21E+07 | 2.53E+04 | 0.41E+04 |
| | 37°C powder | -1.02 | 4.18E+06 | 8.72E+03 | 4.48E+03 |
| 12 Month Stability | Frozen control | N/A | 2.51E+07 | 5.23E+04 | 0.00E+04 |
| | RT powder | -0.81 | 3.87E+06 | 8.06E+03 | 0.81E+03 |
| | 37°C powder | -0.95 | 2.80E+06 | 5.83E+03 | 0.10E+03 |

Stability of Micronized *VitriLife*[®] Powders in Oils

L. Acidophilus in Cottonseed Oil

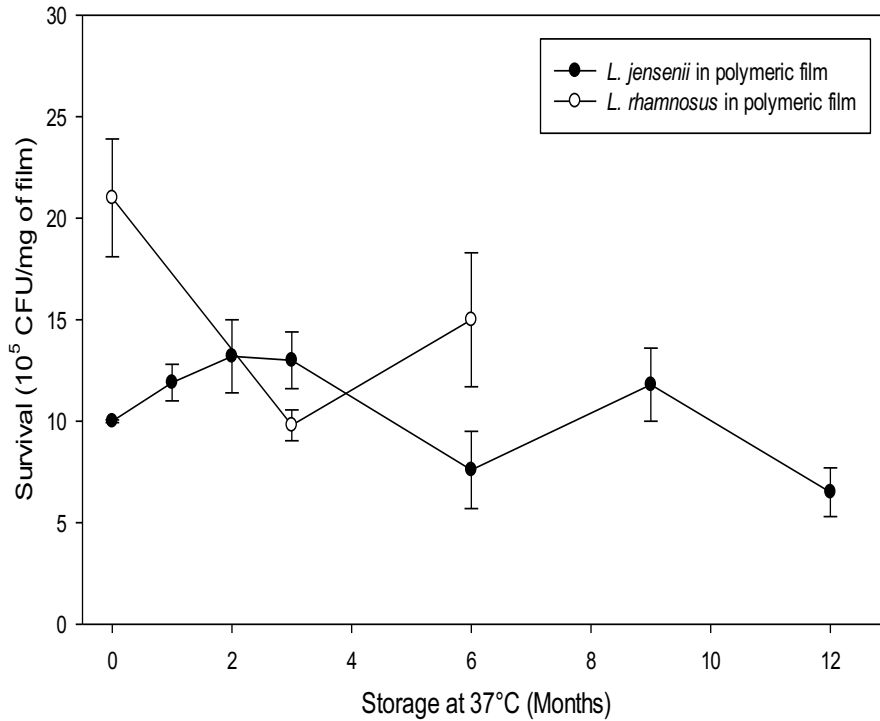


MVA Vaccine in Mineral Oil

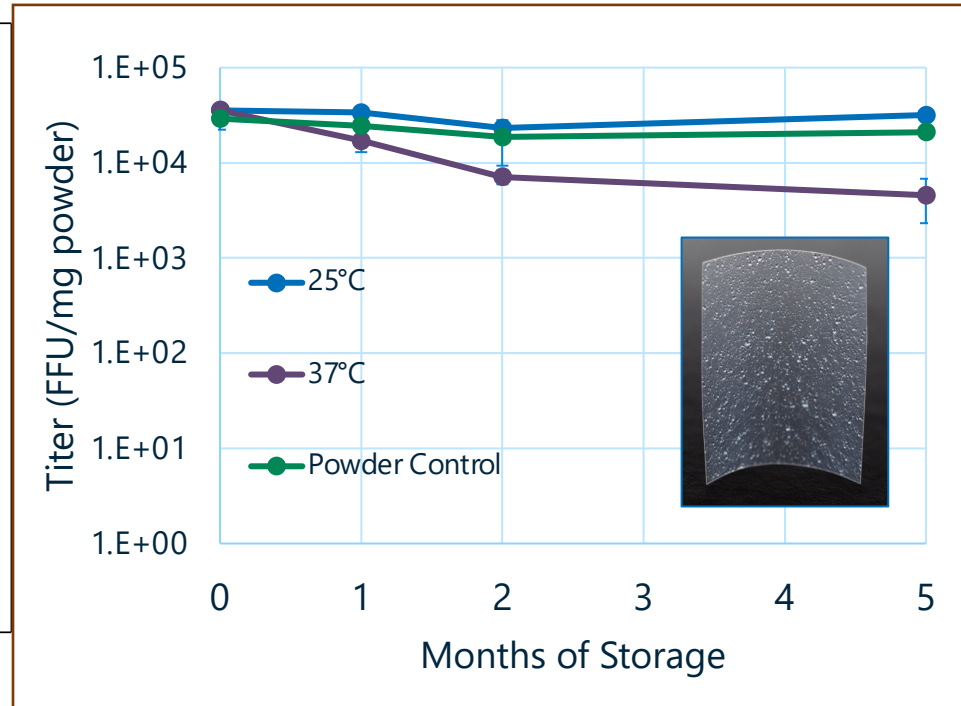


Stability of VitriLife® Powders in HPC Dissolvable Films

Survival of PBV Microbicides in Polymeric Films at 37°C



Stability of PBV Rotavirus LAV in Dissolvable Polymeric Films



VitriLife[®] Inactivation Overview

VitriLife[®] Inactivation (or Sterilization)

Advantages of VitriLife[®]

- ✓ Allows for controllable, targeted damage to replication components
- ✓ Retains membrane structural integrity
- ✓ Live viral vaccines retain high antigenicity after inactivation

Drawbacks of Conventional Inactivation Approaches

Thermal Inactivation:

- ✗ Pervasive damage to cellular components
- ✗ Reduced epithelial adhesion

Chemical Inactivation:

- ✗ Non-specific cross linking leading to the epitope damage
- ✗ Longer treatment times and higher cost
- ✗ Additional processing required to remove residual chemicals

Inactivation by Irradiation in Liquid State:

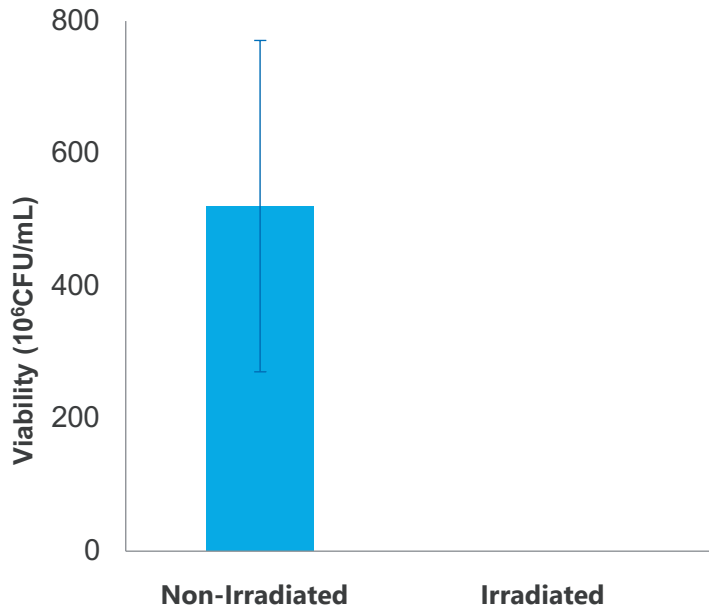
- ✗ Damage of epitopes by free radicals

Production steps of thermostable of UV or electron beam (EB) inactivated vaccines:

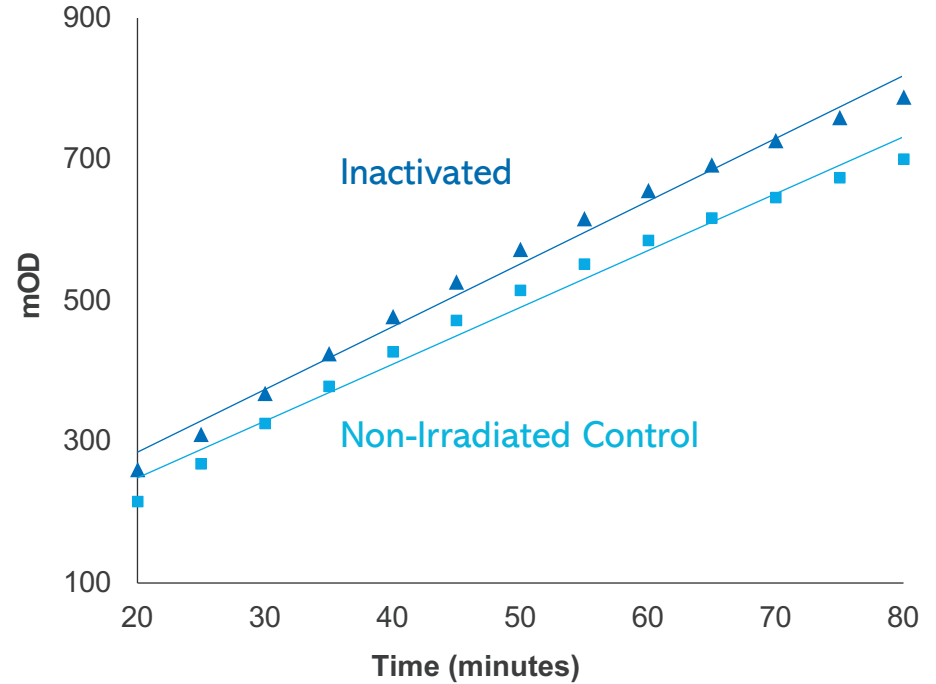


Electron Beam (EB) Inactivated *VitriLife*[®] Bacteria Remained Metabolically Active

12 kGy Dose of EB Irradiation Completely Kills PBV Stabilized E.coli M17

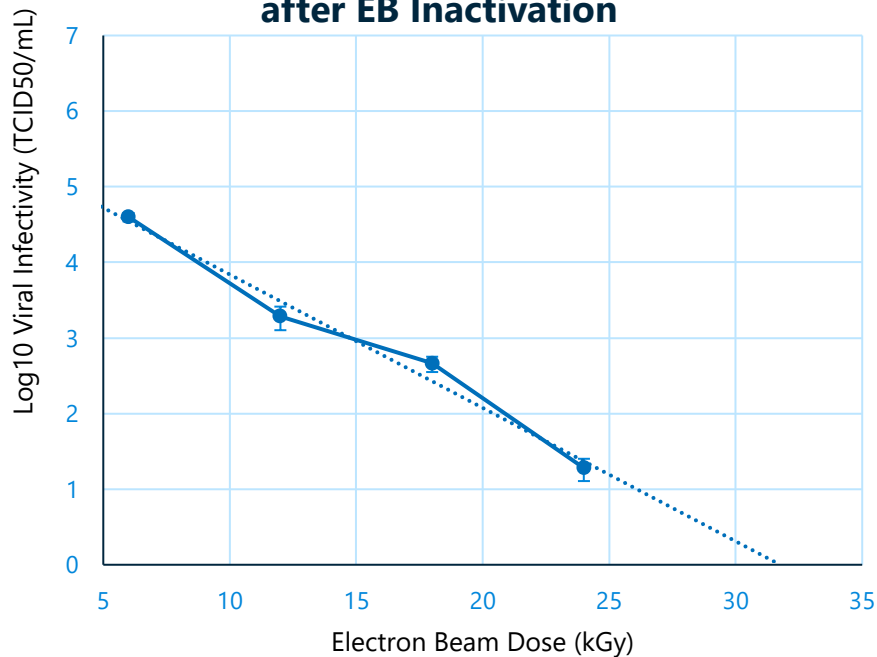


Reducing Metabolic Activity of Both Irradiated and Non-Irradiated *VitriLife*[®] E.coli M17 Remained Equal to 100%



EB Inactivation of *VitriLife*[®] Viruses is a Gentle Process That Does Not Damage Viral Epitopes

VitriLife[®] H3N2 LAIV Infectivity after EB Inactivation

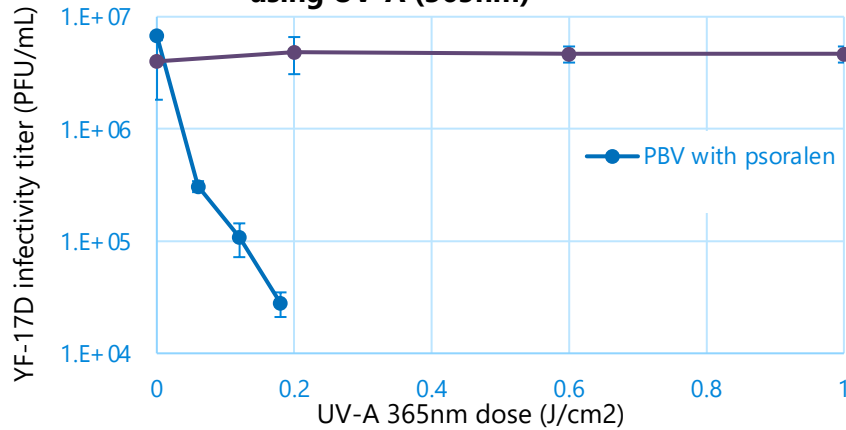


Effect of EB Dose on Infectivity and Antigenicity (HI) of *VitriLife*[®] H3N2 LAIV

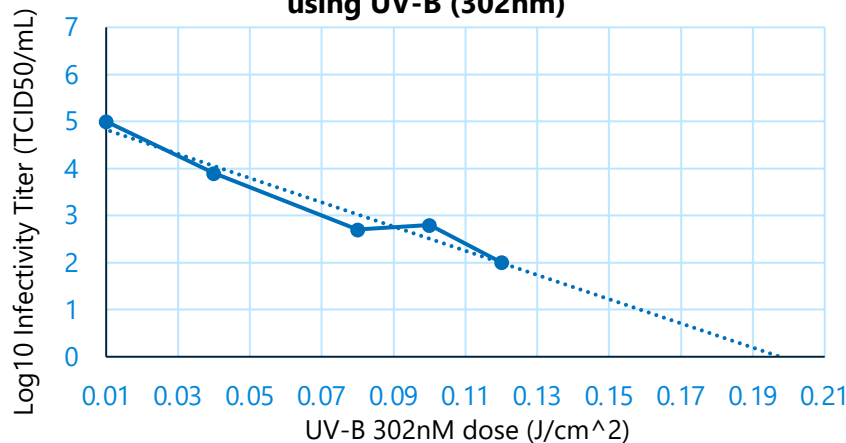
| EB dose | Log Titer Loss | Titer (TCID ₅₀ /mL) | HI Titer |
|---------|----------------|--------------------------------|----------|
| Control | N/A | 6.31E7 | 320 |
| (0kGy) | 0.35 | 2.80 ± 0.36 E7 | 320 |
| 6kGy | 3.20 | 3.97 ± 1.37 E4 | 320 |
| 12kGy | 4.51 | 1.93 ± 0.44 E3 | 320 |
| 18kGy | 5.14 | 4.59 ± 1.5 E2 | 320 |
| 24kGy | >7 | BLD* | 320 |
| 48kGy | NA | BLD* | 285 |

UV Inactivation of *VitriLife*[®] Viruses is a Gentle Process That Does Not Damage Viral Epitopes

Inactivation of *VitriLife*[®] YF-17D LAV using UV-A (365nm)



Inactivation of *VitriLife*[®] H3N2 LAIV using UV-B (302nm)



H3N2 vaccine antigenicity remains undamaged after UV-B inactivation

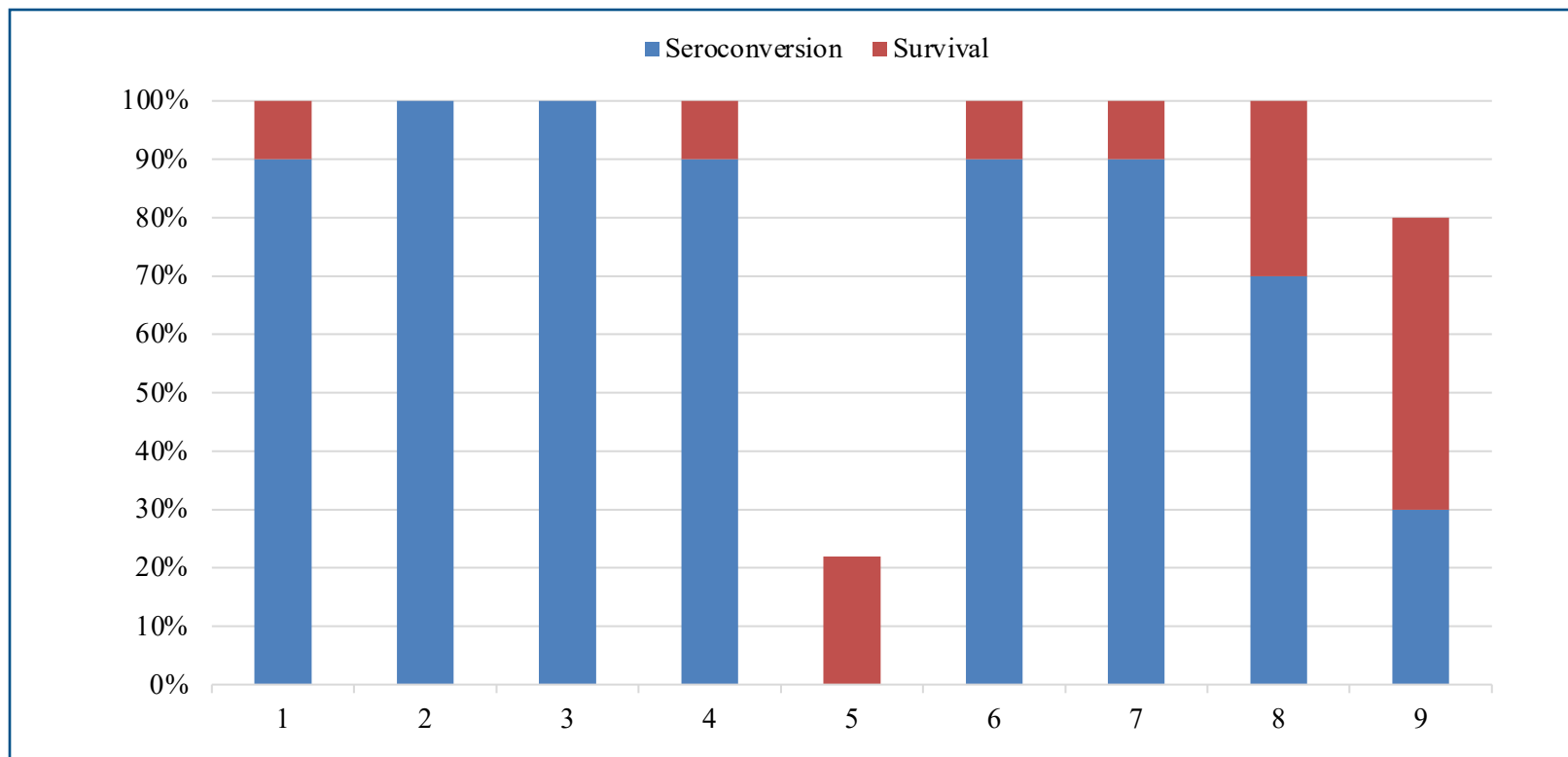
| H3N2 samples | HI titer #1 | HI titer #2 | HI titer #3 | HI titer (geometric mean) |
|---------------------------|-------------|-------------|-------------|---------------------------|
| Frozen control | 320 | 320 | 640 | 403.17 |
| PBV foam control | 320 | 320 | 320 | 320.00 |
| UV 0.04 J/cm ² | 320 | 320 | 320 | 320.00 |
| UV 0.08 J/cm ² | 320 | 320 | 320 | 320.00 |
| UV 0.10 J/cm ² | 320 | 320 | 320 | 320.00 |
| UV 0.12 J/cm ² | 320 | 320 | 320 | 320.00 |
| UV 0.16 J/cm ² | 320 | 320 | 320 | 320.00 |
| UV 0.9 J/cm ² | 320 | 320 | 320 | 320.00 |
| UV 1.8 J/cm ² | 320 | 320 | 320 | 320.00 |

Evaluation of Immunogenicity and Potency of VitriLife[®] Vaccines Using Animal Models

Animal study results were disclosed in the following publications:

- Smith TG, et al. Rabies vaccine preserved by vaporization is thermostable and immunogenic. *Vaccine*. 2015 May;33(19):2203-2206.
- Smith TG, et al. Assessment of the immunogenicity of rabies vaccine preserved by vaporization and delivered to the duodenal mucosa of gray foxes (*Urocyon cinereoargenteus*). *American Journal of Veterinary Research*. 2017;78(6):752-756.
- Hensley C, et al. Thermostable, Dissolvable Buccal Film Rotavirus Vaccine Is Highly Effective in Neonatal Gnotobiotic Pig Challenge Model. *Vaccines*. 2021;9(5):437.
- Luczo JM, et al. Intranasal powder live attenuated influenza vaccine is thermostable, immunogenic, and protective against homologous challenge in ferrets. *NPJ Vaccines*. 2021;6(1).
- Kurup D, et al. Inactivated Rabies Virus-Based Ebola Vaccine Preserved by Vaporization is Heat-Stable and Immunogenic Against Ebola and Protects Against Rabies Challenge. *The Journal of Infectious Diseases*. 2019;220(9):1521-1528.

EB Inactivated *VitriLife*[®] ERA 333 Effectively Protects Mice after IM challenge with Street Rabies Virus

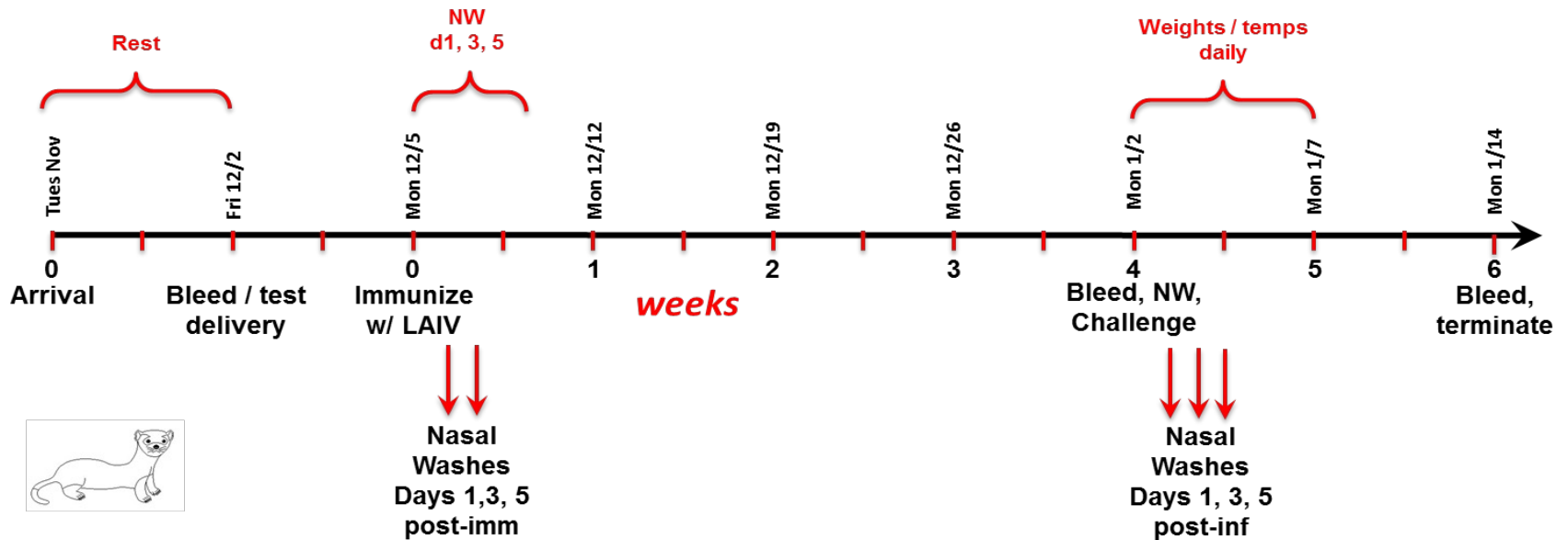


| Dilution Dose (log ₁₀ ffu) | PBV ERAg333 | | | ERAg333 | Placebo | Commer cial | Inactivated PBV ERAg333 | | |
|---|------------------|------------------|------------------|------------------|------------|----------------|----------------------------|------------------|------------------|
| | 10 ⁻¹ | 10 ⁻² | 10 ⁻³ | 10 ⁻² | None | None | None | 10 ⁻¹ | 10 ⁻² |
| | 6.8 | 5.7 | 4.4 | 7.9 | 300 | 620 | 350 | 34 | 2.3 |

*Commercial = RabAvert, Novartis



Immunogenicity and Efficacy of Respiratory VitriLife® H3N2 LAIV in a Ferret Model



Immunization groups

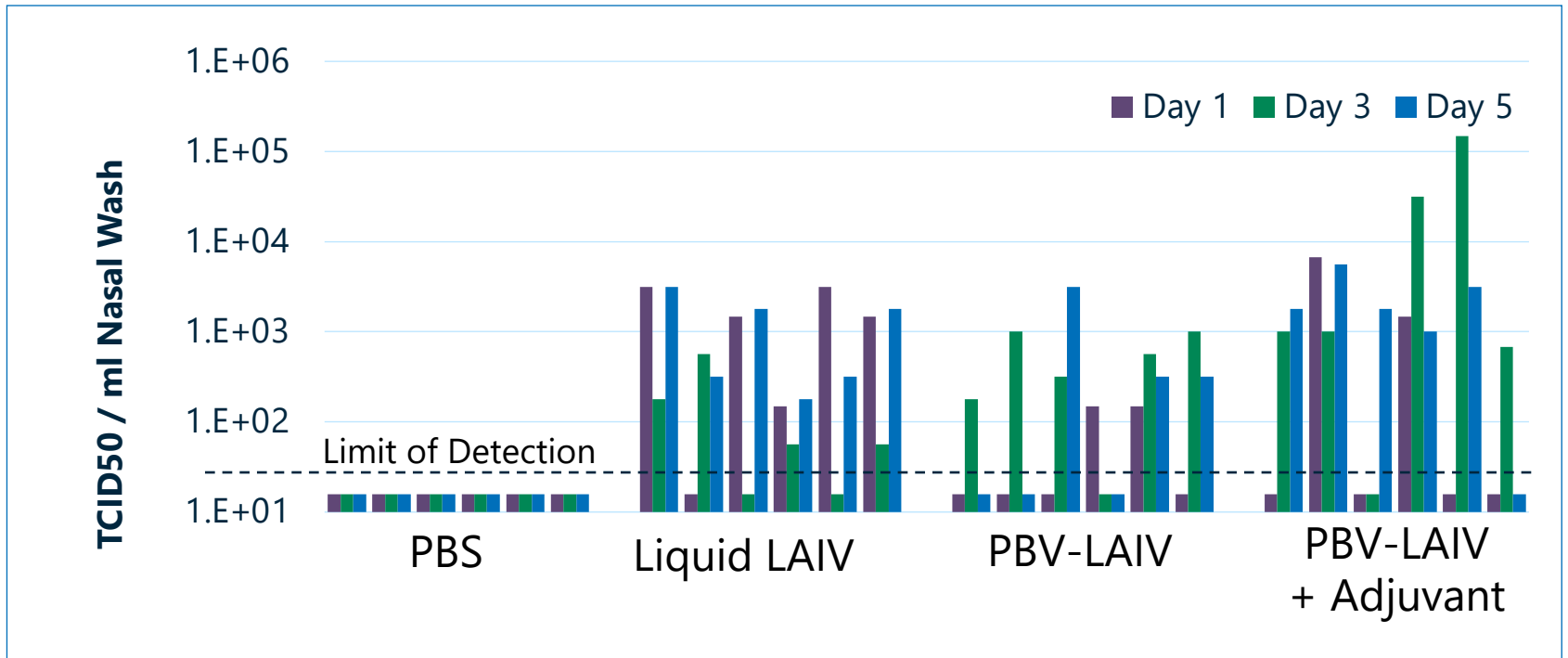
| | |
|--|-------|
| Vehicle/PBS | n = 6 |
| LAIV (-80C liquid stock, 10^7 EID ₅₀) | n = 6 |
| Stabilize LAIV powder (10^7 EID ₅₀) | n = 6 |
| Stabilize LAIV powder + Adj (10^7 EID ₅₀) | n = 6 |

Influenza strains

Vaccine strain: A/17/Texas/2012/30 (H3N2) LAIV (BioDiem)
Stabilization by Universal Stabilization Technologies (UST)
Challenge strain: wildtype A/Texas/50/2012 or similar



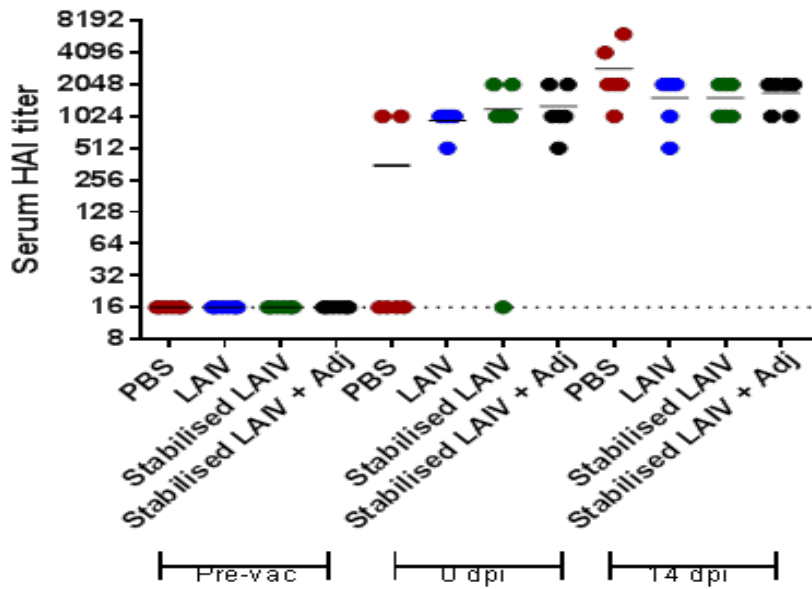
Post-Immunization Nasal Wash H3N2 LAIV Titer



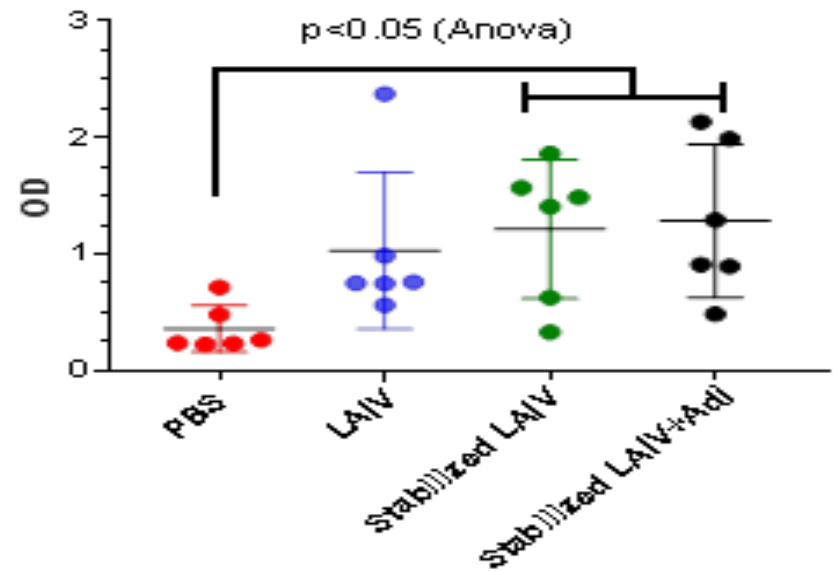


Post-Immunization Nasal Wash Antibodies

Nasal Wash ELISA Titers

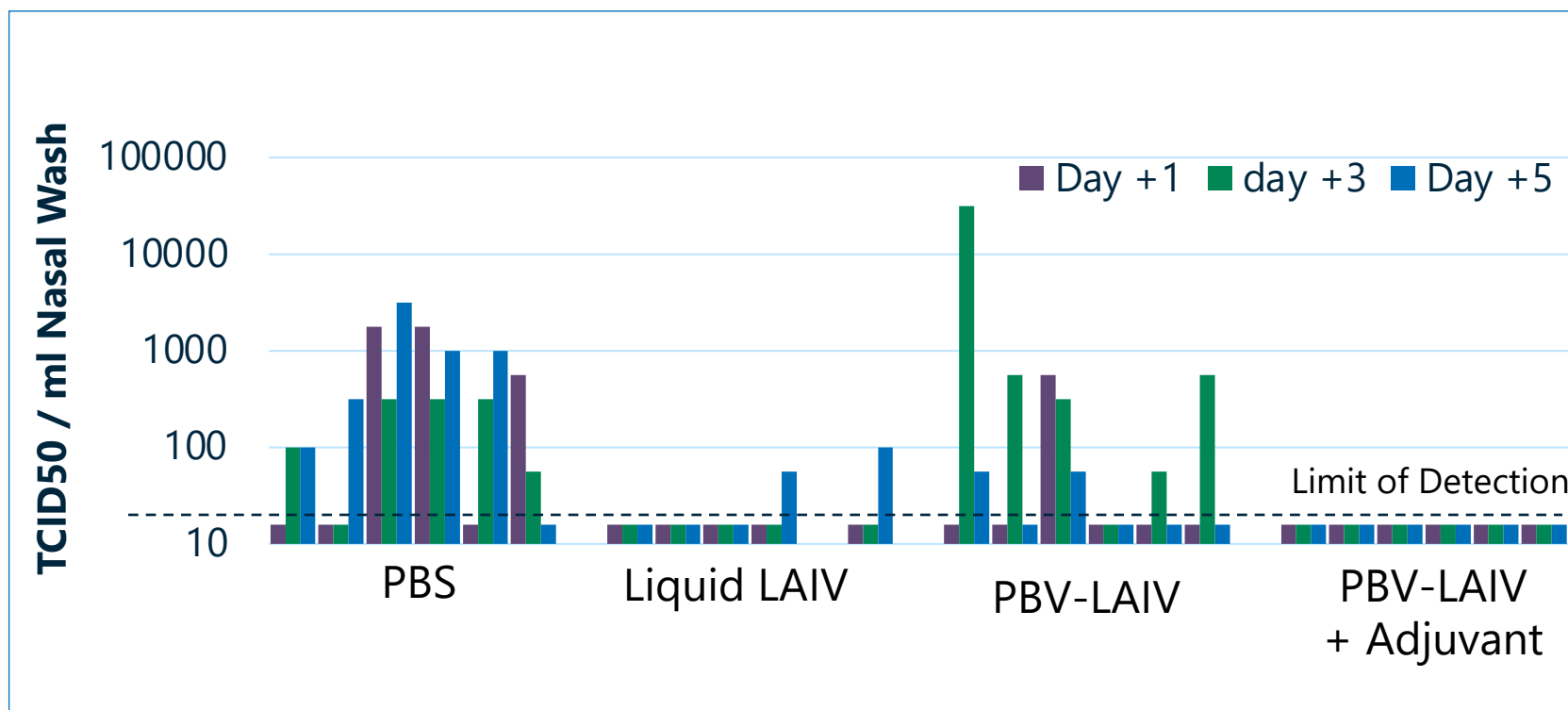


Ferret Ig Response in Nasal Washes Day 1 Post-Challenge (1:4 dilution)

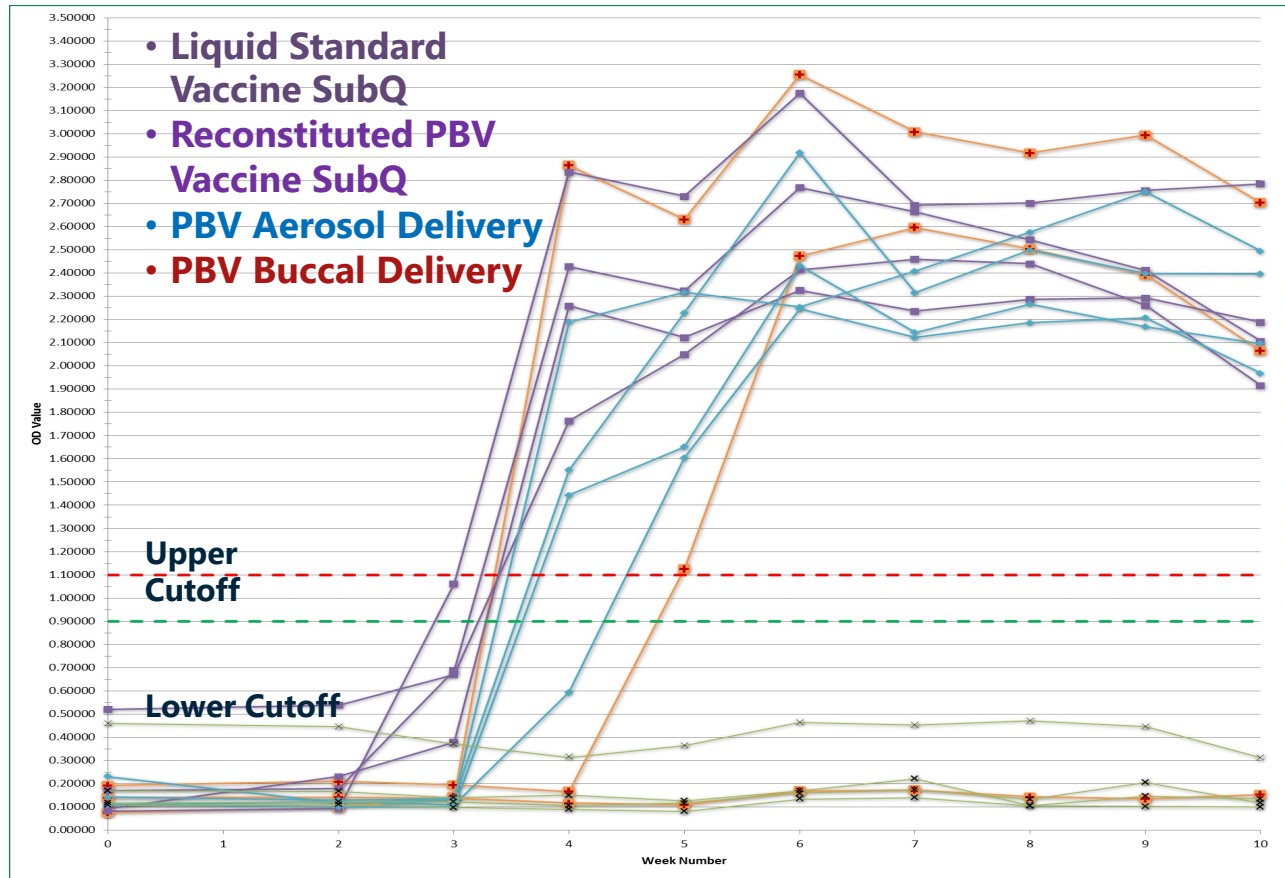




Post-A/TX/50/I2 Challenge Nasal Wash Viral Titers



VitriLife® Measles and Rubella Vaccines Delivered Intranasally in Dry Powder Format were Immunogenic in Non-human Primates



- Intranasal delivery of unadjuvanted aerosolized PBV stabilized dry powders provided high IgG antibody titers after a single dose for rubella vaccine and after a prime + boost regimen for measles vaccine.
- Subcutaneously injected reconstituted PBV measles and rubella vaccines performed as well as the standard liquid vaccines.

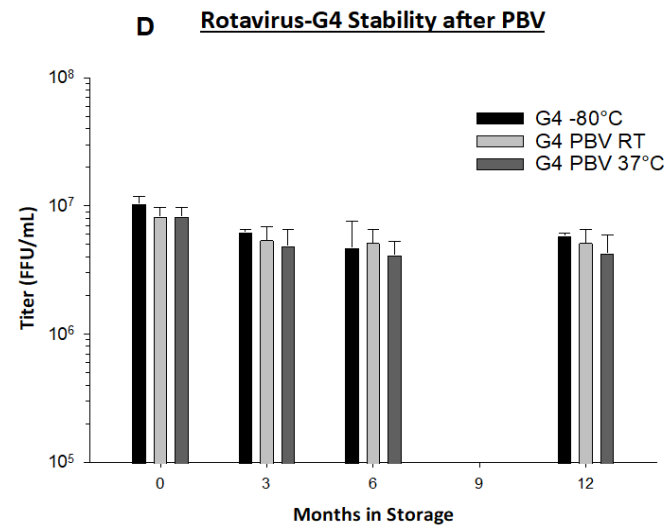
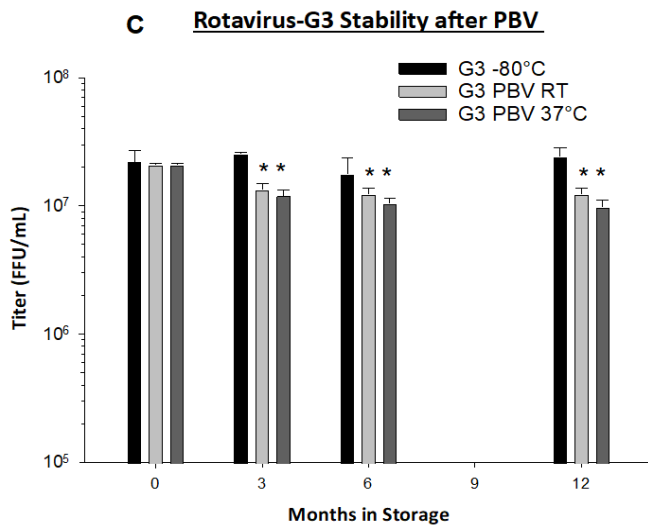
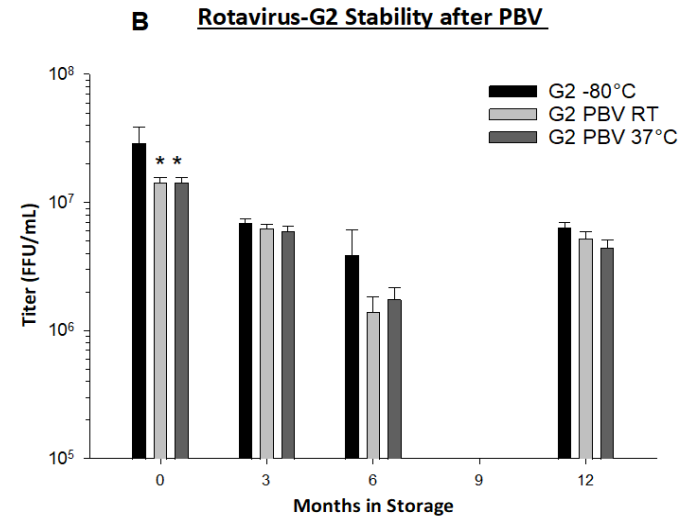
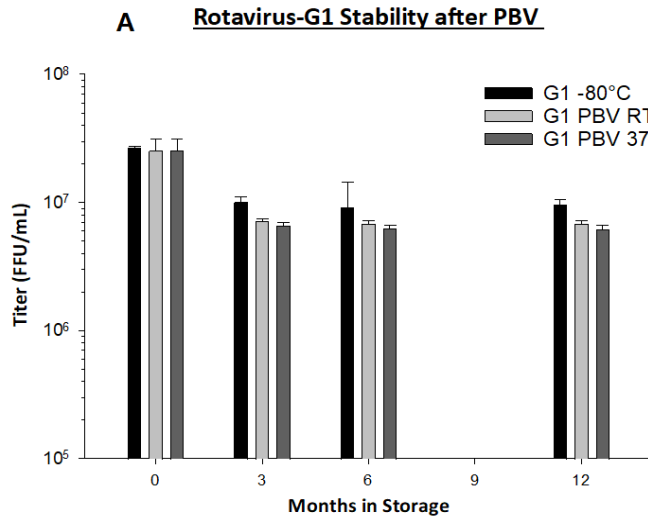
Dissolvable Buccal Films Comprising Micronized *VitriLife*[®] RRV-TV Rotavirus Vaccine and Antacid CaCO₃ powder



Tetravalent rhesus-human reassortant rotavirus vaccine (RRV-TV) includes G1, G2, G3 and G4 mono-reassortant strains.

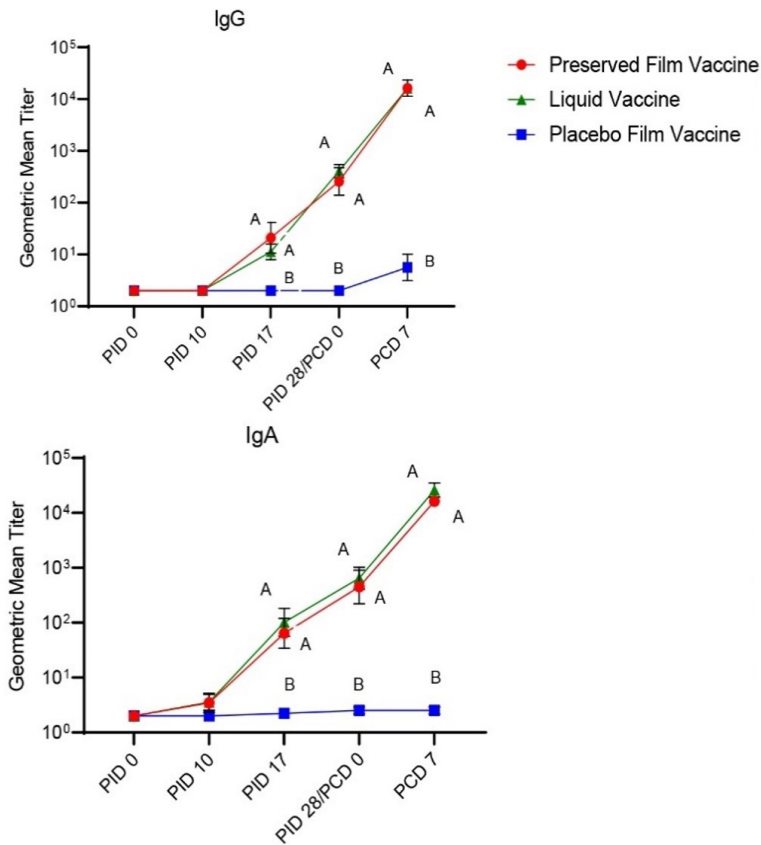
- Initial 14-day toxicity study in rats was performed by Charles River Laboratories. Study results showed no clinical observations of toxicity, effects on body weight or food consumption, changes in hematology or clinical chemistry parameters, or effects on organ weight, and no macroscopic or microscopic findings at any dose level.
- Initial oral mucosal irritation/sensitivity study in hamsters was performed by NAMSA. The right cheek pouch of 10 hamsters was implanted with full human dose film; the left cheek pouch of all animals was implanted with polymethylmethacrylate (PMMA) disks serving as control article. The results showed no macroscopic or microscopic reactions of the RRV-TV film to hamster cheek tissue.

Long-term Stability of PBV Live Rotavirus Vaccines G1 (A), G2 (B), G3 (C) and G4 (D) at Different Storage Temperatures

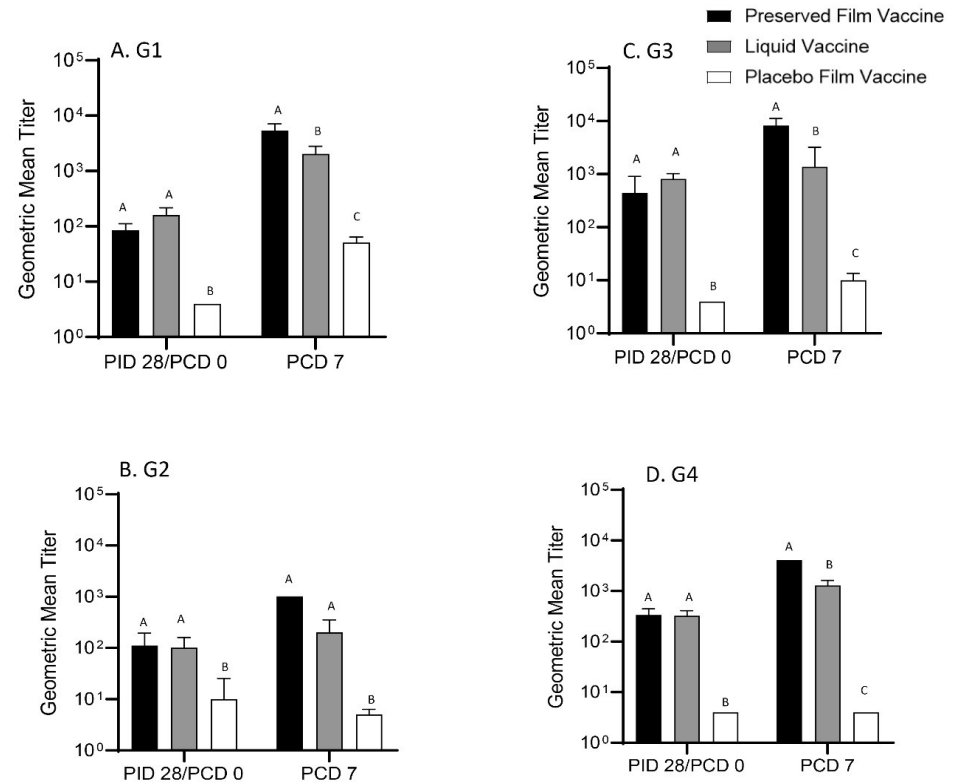


Dissolvable Buccal Film Rotavirus Vaccine Is Highly Effective in Neonatal Gnotobiotic Pig Challenge Model

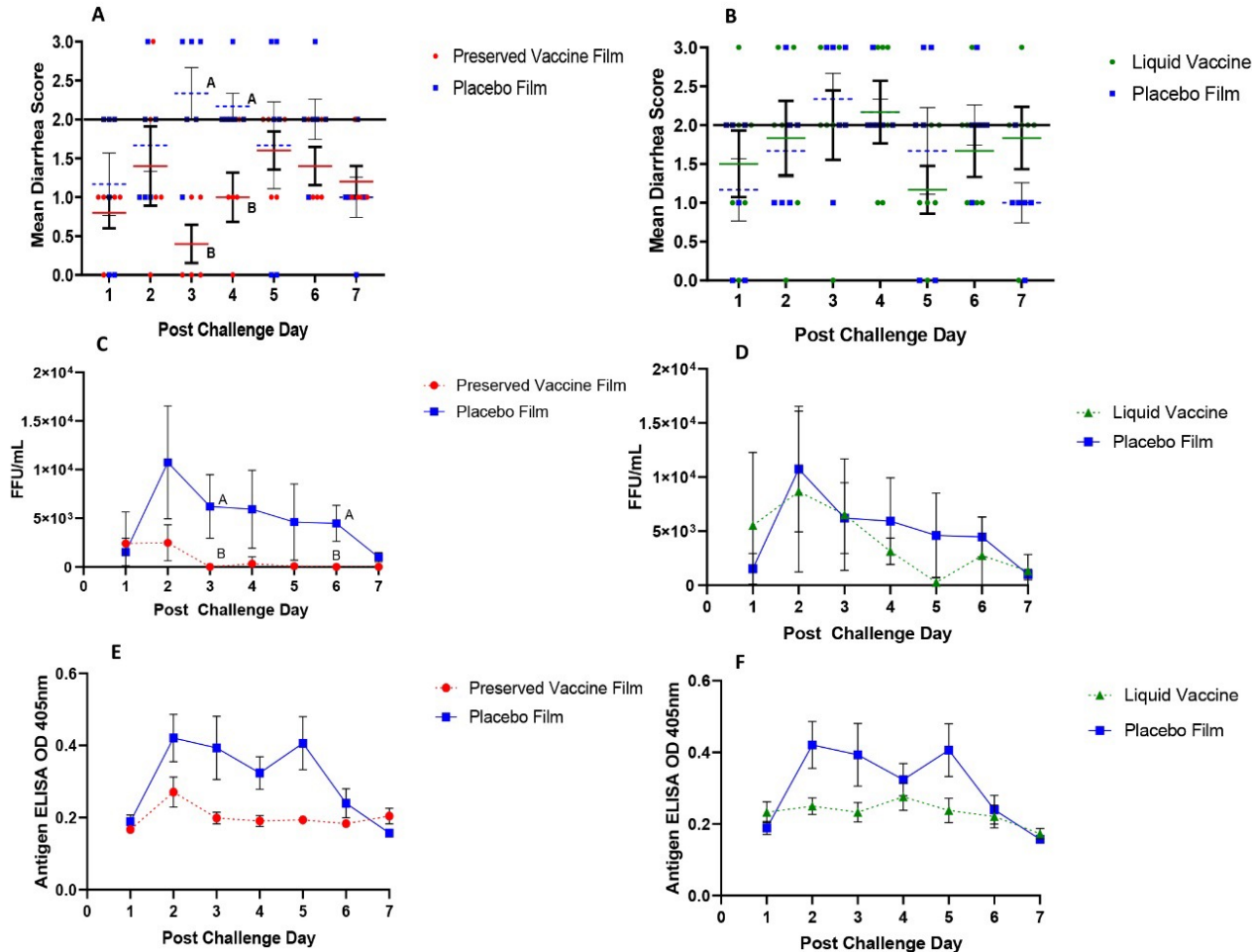
Strong serum IgG and IgA antibody responses were induced by preserved film RRV-TV vaccine.



Significantly higher titers of G1, G3, and G4 rotavirus serum virus neutralization (VN) antibody responses were induced by the preserved film vaccine compared to liquid vaccine at post-challenge day (PCD) 7.



VitriLife® Buccal Film RRV-TV Conferred Strong Protection Against Both Diarrhea and Virus Shedding Upon Challenge with VirHRV



Preserved film-vaccinated pigs showed significantly reduced duration of virus shedding (2.2, 5.7, and 6.8 days, respectively), mean peak titers (~4-fold lower) and area under the curve (AUC) (~6- to 8-fold lower) of virus shedding compared to both placebo and liquid vaccine groups.

Collaborations

NIH and CDC

- Awarded 17 grants and contracts.
- Grants supported studies around AT stabilization of LAVs and microbiome bacteria, mucosal delivery of vaccines and microbiome products.



Military

- Awarded 4 grants.
- Grants supported studies around AT stabilization and delivery of blood components, development of hemostatic and wound healing bandages.



Academic Institutions & Hospitals

- Collaborated with researchers and professors from 10+ academic institutions.



UST is Seeking Validation and Implementation of The *VitriLife*[®] Technology Platform Through:

- Corporate Partnerships
- Strategic Alliances
- Commercial Client Services